

Allergen-specific immunotherapy in children. Consensus document of the Association of Pediatric Allergologists and Immunologists of Russia (positional paper)

REV — обзорная статья

<https://doi.org/10.53529/2500-1175-2023-4-5-30>

Received 15.08.2023

The article is accepted for publication 04.10.2023

Conflict of Interest:

There is no source of funding.

The authors declare a conflict of interest. The authors are members of the editorial board of the journal: Yuri S. Smolkin, Sergey S. Masalsky, Rezeda F. Khakimova, Natalia B. Migacheva.

Yuri S. Smolkin¹, Olga V. Trusova², Zuleikha A. Aliskandieva³, Liudmila Y. Barycheva⁴, Alexey D. Bogomazov⁵, Ksenia A. Bocharova⁶, Yuliya N. Emelina⁷, Andrey V. Kamaev², Inna A. Larkova⁸, Aishat Z. Markhaichuk⁹, Sergey S. Masalskiy¹⁰, Natalia B. Migacheva¹¹, Aleksandr S. Prilutskiy¹², Elena V. Stezhkina¹³, Rezeda M. Fayzullina¹⁴, Rezeda F. Khakimova¹⁵, Ella V. Churyukina¹⁶, Natal'ya V. Shakhova¹⁷, Tatiyana V. Shilova¹⁸

¹ Academy of Postgraduate Education under FSBI FSCC of FMBA of Russia, 91 Volokolamskoe highway, 125371, Moscow, Russia² FSBEI HE I. P. Pavlov SPbSMU MOH Russia, 6–8 Lva Tolstogo st., St. Petersburg, 197022, Russia³ FSBEI HE DSMU MOH Russia, 1 Lenina Sq., Makhachkala, 367012, Russia⁴ FSBEI HE StSMU MOH Russia, 310 Mira st., Stavropol, 355017, Russia⁵ FSBEI HE KSMU MOH Russia, 3 Karla Marksa st., Kursk, 305041, Russia⁶ Belgorod State National Research University, 85 Pobedy st., Belgorod, 308015, Russia⁷ SAHI "Children's State Clinical Hospital No. 9", 51 Reshetskaya st., Sverdlovsk Region, Sverdlovsk Region Center, Yekaterinburg, 620134, Russia⁸ Federal Research Center of Nutrition and Biotechnology, 2/14 Ustinskiy proezd, Moscow, 109240, Russia⁹ GBUZ "Children's Regional Hospital of the Kaliningrad Region", Kaliningrad, 23 D. Donskogo st., 236017, Russia¹⁰ Moscow Medical University "Reaviz", 29 Sokolovo-Meshcherskaya st., 125466, Moscow, Russia¹¹ Samara State Medical University, 137 Samarskaya st., Samara, 443056, Russia¹² FSBEI HE DonSMU of MOH of Russia, 16 Ilyicha av., Donetsk, 283003, Russia¹³ FSBEI HE RyazSMU MOH Russia, 9 Vysokovoltynaya st., Ryazan, 390026, Russia¹⁴ FSBEI HE BSMU MOH Russia, 3 Lenina st., Ufa, 450008, Russia¹⁵ FSBEI HE Kazan SMU MOH Russia, 49 Butlerov st., Kazan, 420012, Russia¹⁶ FSBEI HE RostSMU MOH Russia, 29 Nakhichevanskiy lane, Rostov-on-Don, 344091, Russia¹⁷ FSBEI HE ASMU MOH Russia, 40 Lenin av., Barnaul, 656038, Russia¹⁸ FSBEI HE SUSMU MOH Russia, 64 Vorovsky st., Chelyabinsk, 454092, Russia

Yuri Solomonovich Smolkin — Doc. Sci., Professor of Department of Clinical Immunology and Allergology of Academy of Postgraduate Education under FSBI FSCC of FMBA of Russia, ORCID ID: 0000-0001-7876-6258, smolkin@alerg.ru.

Olga Valerievna Trusova — Cand. Sci., Associate Professor Department of Hospital Therapy with the Course of Allergology and Immunology named after Academician M. V. Chernorutsky with the clinic, FSBEI HE I.P. Pavlov SPbSMU MOH Russia, ORCID ID: 0000-0002-0854-1536, e-mail: o-tru@mail.ru.

Zuleikha Alautdinovna Aliskandieva — Cand. Sci., Associate Professor Department of Faculty and Hospital Pediatrics, FSBEI HE DSMU MOH Russia, ORCID ID: 0009-0005-3836-0428, e-mail: aliskandieva@mail.ru.

Liudmila Yur'evna Barycheva — Dr. Sci., Professor, Head of the Department of Immunology with a course of continuing professional education, FSBEI HE StSMU MOH Russia, ORCID ID: 0000-0002-4069-0566, Stavropol, e-mail: for_liudmila@inbox.ru.

Alexey Dmitrievich Bogomazov — Cand. Sci., Associate Professor Department of Pediatrics, FSBEI HE KSMU MOH Russia, ORCID ID: 0000-0002-4636-1819, e-mail: bogomazov71@mail.ru.

Ksenia Aleksandrovna Bocharova — Cand. Sci., Associate Professor, Associate Professor of Department of Family Medicine of the Medical Institute, Belgorod State National Research University, ORCID ID: 0000-0001-5540-924X, e-mail: doctor.bocharova@mail.ru.

For correspondence:

Yuri Solomonovich Smolkin, Professor of Department of Clinical Immunology and Allergology Academy of Postgraduate Education under FSBI FSCC of FMBA of Russia.

Address: 91 Volokolamskoe shosse, Moscow, 125371, Russia.

E-mail: smolkin@alerg.ru.

Для корреспонденции:

Смолкин Юрий Соломонович, профессор кафедры клинической иммунологии и аллергологии Академии постдипломного образования ФГБУ ФНКЦ ФМБА России.

Адрес: 125371, Россия, г. Москва, Волоколамское ш., 91.

E-mail: smolkin@alerg.ru.

Yuliya Nikolaevna Emelina — Cand. Sci., allergist-immunologist, SAHI “Children’s State Clinical Hospital No. 9”, ORCID ID: 0009-0005-3836-0428, e-mail: eyun75@mail.ru.

Andrey Vyacheslavovich Kamaev — Cand. Sci., Associate Professor, Associate Professor Department of General Medical Practice, FSBEI HE I.P. Pavlov SPbSMU MOH Russia, ORCID ID: 0000-0001-9654-3429, e-mail: andykkam@mail.ru.

Inna Anatolevna Larkova — Cand. Sci., Senior researcher of Allergy Department, Federal Research Center of Nutrition and Biotechnology, ORCID ID: 0000-0001-7640-0754, e-mail: inna_larkova@mail.ru.

Aishat Ziyabutdinovna Markhaichuk — allergologist-immunologist, Consultative and Diagnostic Center for Children, Children’s Regional Hospital of the Kaliningrad Region, ORCID ID: 0009-0000-4513-2208, e-mail: ayshat.90@rambler.ru.

Sergey Sergeevich Masalskiy — Cand. Sci., Associate Professor of the Department of Obstetrics, Gynecology and Pediatrics, Moscow Medical University “Reaviz”, ORCID ID: 0000-0002-2048-5709, e-mail: masalsky@live.com.

Natalia Begievna Migacheva — Dr. Sci., Associate Professor, Head of Department of Pediatrics, Samara State Medical University, ORCID ID: 0000-0003-0941-9871, Samara, e-mail: nbmigacheva@gmail.com.

Aleksandr Sergeevich Prilutskiy — Dr. Sci., Professor of Department of Microbiology, Virology, Immunology and Allergology, FSBEI HE DonSMU of MOH of Russia, ORCID ID: 0000-0003-1409-504X, Donetsk, e-mail: aspr@mail.ru.

Elena Viktorovna Stezhkina — Cand. Sci., Associate Professor of the Department of Faculty and Polyclinic Pediatrics with the Course of Pediatrics FDPO, FSBEI HE RyazSMU MOH Russia, ORCID ID: 0000-0002-1806-0787, Ryazan, e-mail: polus1972@yandex.ru.

Rezeda Mansafonva Fayzullina — Dr. Sci., Professor of the Department of Faculty Pediatrics with Courses in Pediatrics, Neonatology and Simulation Center IDPO, FSBEI HE BSMU MOH Russia, ORCID ID: 0000-0002-9001-1437, Ufa, e-mail: fayzullina@yandex.ru.

Rezeda Fidailovna Khakimova — Dr. Sci., Professor of the Department of Clinical Immunology and Allergology, FSBEI HE Kazan SMU MOH Russia, ORCID ID: 0000-0003-0754-9605, Kazan, e-mail: khakimova@yandex.ru.

Ella Vitalievna Churyukina — Cand. Sci., Associate Professor of the Department of Clinical Immunology, Allergology and laboratory diagnostics, FSBEI HE RostSMU MOH Russia, ORCID ID: 0000-0001-6407-6117, Rostov-on-Don, e-mail: echuryukina@mail.ru.

Natal’ya Victorovna Shakhova — Dr. Sci., Associate Professor, Head of the Department of Hospital Pediatrics with a Course of Additional Professional Education, FSBEI HE ASMU MOH Russia, ORCID ID: 0000-0002-7143-8259, Barnaul, e-mail: natalia.shakhova@mail.ru.

Tatiana Vasilevna Shilova — Associate Professor Department of Hospital Pediatrics, Clinical Immunology and Allergology FSBEI HE SUSMU MOH Russia, e-mail: tanya920477@mail.ru.

Annotation

The Document contains fundamental provisions regarding allergen -specific immunotherapy in children. International clinical recommendations on the methodology were used, adapted for use in real practice. In the absence of international recommendations, the authors present the consensus opinion of the project participants, based on data from clinical studies in this area. At the moment, we bring to your attention a position paper on allergen-specific immunotherapy in children, created by experts from the Association of Pediatric Allergologists and Immunologists of Russia (APAIR) based on the 2021 consensus document, with the necessary data updates.

Keywords: allergen-specific immunotherapy, children, consensus document.

For citation: Smolkin YS, Trusova OV, Aliskandieva ZA, Barycheva LY, Bogomazov AD, Bocharova KA, Emelina YN, Kamaev AV, Larkova IA, Markhaichuk AZ, Masalskiy SS, Migacheva NB, Prilutskiy AS, Stezhkina EV, Fayzullina RM, Khakimova RF, Churyukina EV, Shakhova NV, Shilova TV. Allergen-specific immunotherapy in children. Consensus document of the Association of Pediatric Allergologists and Immunologists of Russia (positional paper). *Allergology and Immunology in Pediatrics*. 2023; 3: 5–30. <https://doi.org/10.53529/2500-1175-2023-4-5-30>

Аллерген-специфическая иммунотерапия у детей. Согласительный документ Ассоциации детских аллергологов и иммунологов России (позиционная статья)

<https://doi.org/10.53529/2500-1175-2023-4-5-30>

Статья поступила 15.08.2023

Статья принята в печать 04.10.2023

УДК 616-035

Конфликт интересов:

Источник финансирования отсутствует.

Авторы заявляют о конфликте интересов. Авторы входят в редакционную коллегию журнала: Смолкин Ю. С., Масальский С. С., Хакимова Р. Ф., Мигачева Н. Б., Чурюкина Э. В.

Статья прошла двойное слепое внешнее рецензирование.

Смолкин Ю. С.¹, Трусова О. В.², Алискандиева З. А.³, Барычева Л. Ю.⁴, Богомазов А. Д.⁵, Бочарова К. А.⁶, Емелина Ю. Н.⁷, Камаев А. В.², Ларькова И. А.⁸, Мархайчук А. З.⁹, Масальский С. С.¹⁰, Мигачева Н. Б.¹¹, Прилуцкий А. С.¹², Стежкина Е. В.¹³, Файзуллина Р. М.¹⁴, Хакимова Р. Ф.¹⁵, Чурюкина Э. В.¹⁶, Шахова Н. В.¹⁷, Шилова Т. В.¹⁸

- ¹ Академия постдипломного образования ФГБУ ФНКЦ ФМБА России, 125371, г. Москва, Волоколамское ш., д. 91, Россия
- ² ФГБОУ ВО ПСПбГМУ им. И. П. Павлова Минздрава России, 197022, г. Санкт-Петербург, ул. Льва Толстого, д. 6–8, Россия
- ³ ФГБОУ ВО ДГМУ Минздрава России, 367012, г. Махачкала, пл. им. Ленина, д. 1, Россия
- ⁴ ФГБОУ ВО СтГМУ Минздрава России, 355017, г. Ставрополь, ул. Мира, д. 310, Россия
- ⁵ ФГБОУ ВО КГМУ Минздрава России, 305041, г. Курск, ул. Карла Маркса, д. 3, Россия
- ⁶ ФГАОУ ВО «Белгородский государственный национальный исследовательский университет», 308015, г. Белгород, ул. Победы, д. 85, Россия
- ⁷ ГАУЗ ДГКБ № 9, 620134, г. Екатеринбург, ул. Решетская, д. 51, Россия
- ⁸ ФГБУН «ФИЦ питания и биотехнологии», 109240, г. Москва, Устьинский пр., д. 2/14, Россия
- ⁹ ГБУЗ «Детская областная больница Калининградской области», 236017, г. Калининград, ул. Д. Донского, д. 23, Россия
- ¹⁰ Московский медицинский университет «Реавиз», 125466, г. Москва, ул. Соколово-Мещерская, д. 29, Россия
- ¹¹ ФГБОУ ВО СамГМУ Минздрава России, 443056, г. Самара, ул. Самарская, д. 137, Россия
- ¹² ФГБОУ ВО ДонГМУ Минздрава России, 283003, г. Донецк, пр-т Ильича, д. 16, Россия
- ¹³ ФГБОУ ВО РязГМУ Минздрава России, 390026, г. Рязань, ул. Высоковольтная, д. 9, Россия
- ¹⁴ ФГБОУ ВО БГМУ Минздрава России, 450008, г. Уфа, ул. Ленина, д. 3, Россия
- ¹⁵ ФГБОУ ВО «Казанский государственный медицинский университет» Минздрава России, 420012, г. Казань, ул. Бутлерова, д. 49, Россия
- ¹⁶ ФГБОУ ВО РостГМУ Минздрава России, 344091, г. Ростов-на-Дону, пер. Нахичеванский, д. 29, Россия
- ¹⁷ ФГБОУ ВО АГМУ Минздрава, 656038, г. Барнаул, пр-т Ленина, д. 40, Россия
- ¹⁸ ФГБОУ ВО ЮУГМУ Минздрава России, 454092, г. Челябинск, ул. Воровского, д. 64, Россия

Смолкин Юрий Соломонович — д. м. н., профессор кафедры клинической иммунологии и аллергологии Академии постдипломного образования ФГБУ ФНКЦ ФМБА России, ORCID ID: 0000-0001-7876-6258, e-mail: smolkin@alerg.ru.

Трусова Ольга Валерьевна — к. м. н., доцент кафедры терапии госпитальной с курсом аллергологии и иммунологии им. ак. М. В. Черноруцкого с клиникой ФГБОУ ВО ПСПбГМУ им. И. П. Павлова Минздрава России, ORCID ID: 0000-0002-0854-1536, e-mail: o-tru@mail.ru.

Алискандиева Зулейха Алаутдиновна — к. м. н., доцент кафедры факультетской и госпитальной педиатрии ФГБОУ ВО ДГМУ Минздрава России, ORCID ID: 0009-0005-3836-0428, e-mail: aliskandieva@mail.ru.

Барычева Людмила Юрьевна — д. м. н., профессор, заведующая кафедрой иммунологии с курсом ДПО ФГБОУ ВО СтГМУ Минздрава России, ORCID ID: 0000-0002-4069-0566, e-mail: for_ludmila@inbox.ru.

Богомазов Алексей Дмитриевич — к. м. н., доцент кафедры педиатрии ФГБОУ ВО КГМУ Минздрава России, ORCID ID: 0000-0002-4636-1819, e-mail: bogomazov71@mail.ru.

Бочарова Ксения Александровна — к. м. н., доцент, доцент кафедры семейной медицины медицинского института ФГАОУ ВО «Белгородский государственный национальный исследовательский университет», ORCID ID: 0000-0001-5540-924X, e-mail: doctor.bocharova@mail.ru.

Емелина Юлия Николаевна — к. м. н., врач аллерголог-иммунолог, ГАУЗ «Детская государственная клиническая больница № 9», ORCID ID: 0009-0005-3836-0428, e-mail: eyun75@mail.ru.

Камаев Андрей Вячеславович — к. м. н., доцент, доцент кафедры общей врачебной практики (семейной медицины) ФГБОУ ВО ПСПбГМУ им. И. П. Павлова Минздрава России, ORCID ID: 0000-0001-9654-3429, e-mail: andykham@mail.ru.

Ларькова Инна Анатольевна — к. м. н., старший научный сотрудник отделения аллергологии ФГБУН «ФИЦ питания и биотехнологии», ORCID ID: 0000-0001-7640-0754, e-mail: inna_larkova@mail.ru.

Мархайчук Айшат Зиябутдиновна — врач аллерголог-иммунолог Консультативно-диагностического центра для детей ГБУЗ «Детская областная больница Калининградской области», ORCID ID: 0009-0000-4513-2208, e-mail: ayshat.90@rambler.ru

Масальский Сергей Сергеевич — к. м. н., доцент кафедры акушерства, гинекологии и педиатрии Медицинского университета «Реавиз», ORCID ID: 0000-0002-2048-5709, e-mail: masalsky@live.com.

Мигачева Наталья Бегиевна — д. м. н., доцент, заведующая кафедрой педиатрии ИПО ФГБОУ ВО СамГМУ Минздрава России, ORCID ID: 0000-0003-0941-9871, e-mail: nbmigacheva@gmail.com.

Прилуцкий Александр Сергеевич — д. м. н., профессор кафедры микробиологии, вирусологии, иммунологии и аллергологии ФГБОУ ВО ДонГМУ Минздрава России, ORCID ID: 0000-0003-1409-504X, e-mail: aspr@mail.ru.

Стежкина Елена Викторовна — к. м. н., доцент кафедры факультетской и поликлинической педиатрии с курсом педиатрии ФДПО ФГБОУ ВО РязГМУ Минздрава России, ORCID ID: 0000-0002-1806-0787, e-mail: polus1972@yandex.ru.

Файзуллина Резеда Мансафовна — д. м. н., профессор кафедры факультетской педиатрии с курсами педиатрии, неонатологии и симуляционным центром ИДПО ФГБОУ ВО БГМУ Минздрава России, ORCID ID: 0000-0002-9001-1437, e-mail: fayzullina@yandex.ru.

Хакимова Резеда Фидаиловна — д. м. н., профессор кафедры клинической иммунологии с аллергологией ФГБОУ ВО Казанский ГМУ Минздрава России, ORCID ID: 0000-0003-0754-9605, e-mail: khakimova@yandex.ru.

Чурыкина Элла Витальевна — к.м.н., доцент, начальник отдела аллергических и аутоиммунных заболеваний в педиатрии НИИАП ФГБОУ ВО Ростовский государственный медицинский университет Минздрава России; доцент кафедры клинической иммунологии, аллергологии и лабораторной диагностики ФПК и ППС ФГБОУ ВО Кубанский государственный медицинский университет Минздрава России. Врач аллерголог-иммунолог высшей категории, ORCID ID: 0000-0001-6407-6117, e-mail: echuryukina@mail.ru.

Шахова Наталья Викторовна — д.м.н., доцент, заведующий кафедрой госпитальной педиатрии с курсом ДПО ФГБОУ ВО АГМУ Минздрава, ORCID ID: 0000-0002-7143-8259, e-mail: natalia.shakhova@mail.ru.

Шилова Татьяна Васильевна — к.м.н., доцент кафедры госпитальной педиатрии, клинической иммунологии и аллергологии ФГБОУ ВО ЮУГМУ Минздрава России, ORCID ID: 0000-0001-9826-9654, e-mail: tanya920477@mail.ru.

Аннотация

В Документе содержатся принципиальные положения, касающиеся проведения аллерген-специфической иммунотерапии у детей. Используются международные клинические рекомендации по методике, адаптированные для применения в условиях реальной практики. В случае отсутствия международных рекомендаций авторами представлено консенсусное мнение участников проекта, основанное на данных клинических исследований. В настоящий момент мы предлагаем вашему вниманию позиционный документ по вопросам аллерген-специфической иммунотерапии у детей, созданный экспертами Ассоциации детских аллергологов и иммунологов России (АДАИР) на основе согласительного документа 2021 г., с необходимыми обновлениями данных.

Ключевые слова: аллерген-специфическая иммунотерапия, дети, согласительный документ.

Для цитирования: Смолкин ЮС, Трусова ОВ, Алискандиева ЗА, Барычева ЛЮ, Богомазов АД, Бочарова КА, Емелина ЮН, Камаев АВ, Ларькова ИА, Мархайчук АЗ, Масальский СС, Мигачева НБ, Прилуцкий АС, Стежкина ЕВ, Файзуллина РМ, Хакимова РФ, Чурыкина ЭВ, Шахова НВ, Шилова ТВ. Аллерген-специфическая иммунотерапия у детей. Согласительный документ Ассоциации детских аллергологов и иммунологов России (позиционная статья). *Аллергология и иммунология в педиатрии*. 2023; 4: 5–30. <https://doi.org/10.53529/2500-1175-2023-4-5-30>

Allergen-specific immunotherapy (ASIT) was offered more than 100 years ago and showed its effectiveness in the therapy of diseases, mediated by specific class E antibodies (IgE). The evidence base of ASIT efficiency and safety is supported by the results of current research of the highest level and continues to be replenished with. At least one third of reports are dedicated to various aspects of ASIT use at congresses of European and American Academies of Allergy. ASIT is the only method, which can change the course of allergic disease towards remission. Nevertheless, a very small percentage of patients with allergic disease of IgE-dependent type and confirmed sensitization to pollen, household and other kinds of allergens receive this type of treatment.

The benefit of the national allergology is the existence of the school, founded by Academician Andrey Dmitrievich Ado. Our teachers and mentors managed to achieve major breakthrough in the field of allergology and immunology in the 70–80s of XX century. Allergy service was built at a high level in the Soviet Union, and ASIT was applied to a larger number of patients. Nowadays the slow advent of modern allergy vaccine in our market predetermines limited therapeutic options for the allergist-immunologist, not to mention that the process for the creation of own modern allergen drugs is just getting underway in our country.

ASIT HISTORY

ASIT history is 110-year-old way from empirical knowledge to evidence-based medicine. Leonard Noon is called ASIT father, who successfully tested pre-season subcutaneous injections of pollen extracts in increasing doses to treat hay fever, calling the method “preventive vaccination” [1]. R. Cook suggested using extracts of animal dander, food and insects for treatment as well as developed a method used to standardize allergens extracts with the Kejldahl method to determine PNU (protein nitrogen units), which was widely used in the world up to the 1980s, and it is applied in Russia even now [2]. In his works D. Freeman offered expedited ASIT schemes and described adverse local and systematic reactions [3]. Since the beginning of XX century, apart from the classical method of subcutaneous injection, intradermal, intralymphatic, oral, sublingual, nasal, conjunctival methods of ASIT have been actively investigated [4].

Allergen extracts were replaced by allergoids in 1970, improving the efficiency and safety of ASIT. Formaldehyde, glutaraldehyde and polyethylene glycol were used for allergen chemical modification. Physical modification of allergens was carried out using aluminum hydroxide, calcium salts and L-tyrosine. A new milestone in allergology was the creation of recombinant allergens in the late 1980s.

The great merit in forming and developing ASIT method belongs to the USSR allergists. Under the leadership of academician A. D. Ado the production of therapeutic and diagnostic allergens was organized and ASIT methods were developed, being used today [5, 6]. Led by the corresponding member of RAS, Professor I. I. Balabolkin, the school of Pediatric Allergology was established, applying ASIT with non-infectious and infectious allergens in patients with bronchial asthma (BA), allergic rhinitis (AR), atopic dermatitis (AtD) as well as the scheme of sublingual administration of water-salt allergen extracts was developed. The works of Dr. of Sci Yu. S. Smolkin in the 90s of the XX century showed the comparative efficacy of subcutaneous and sublingual methods of ASIT. In current years the evidence base on ASIT efficacy with various allergens is updated, the mechanisms of the method acting and interesting aspects of ASIT effect are investigated, first of all, a disease-modifying prolonged effect and possibility to prevent progression in patients with BA.

MECHANISMS OF ALLERGEN-SPECIFIC IMMUNOTHERAPY

1. ASIT is the method of treating allergic diseases, aimed at building immunological tolerance to certain allergens.
2. The multifaceted mechanism of ASIT action involves humoral (production of blocking antibodies) and cellular (stimulation of cell proliferation and differentiation of regulatory cell subpopulations) mechanisms of the immune response.

ASIT acts as humoral (production of blocking antibodies) and cellular (stimulation of cell proliferation and differentiation of regulatory cell subpopulations) mechanisms of the immune response that provides for the suppression of allergic inflammation in the tissues of target organs. In the first weeks of using ASIT there is a transient increase and then decrease in the level of specific IgE blood serum without any connection with symptoms of the disease. In 4–6 weeks from the start of treatment there is an increase in concentrations of specific blocking antibodies of Class IgA and IgG (subclasses 1–4) [7]; it results in oppression of effector cells (mast cells, basophiles, eosinophils) [8]. Due to deactivation of effector cells, there is a reduction in the secretion of a wide range of mediators and cytokines that prevents further stimulation of type 2 T-helpers, eosinophils,

etc. ASIT mechanisms affect both phases of the allergic response — early and late. Tissue specific response to allergen is inhibited, in particular by increasing sensitivity threshold to histamine [9].

The intake of therapeutic allergen suppresses cells, polarizing the immune response towards allergic inflammation, in the part of innate immunity (type 2 innate lymphoid cells (ILC2)) as well as in the adaptive one (Th2 and Th17 cells) and reduces their production of T2 cytokines [8, 10]. ASIT stimulates tolerogenic subpopulations of dendritic and congenital lymphoid cells and also induces allergen-specific regulatory T cells (Treg) and B cells (Breg), whose combined work is most important for creating immunological tolerance, realized through tolerogenic cytokines (interleukin 10 (IL-10)), interleukin 35 (IL-35), transforming growth factor beta (TGF- β) and etc.) [7, 8, 11]. Treg perform early suppression of effector cells, suppression of inflammatory dendritic cells, ILC2, Th1, Th2, Th17, contribute to Breg formation.

ASIT efficiency (formation of tolerance) depends both on endogenous and exogenous factors; the type, method and amount of allergen administered have the greatest impact [12].

IMMUNOLOGICAL FEATURES OF INDUCED TOLERANCE IN CHILDREN

1. ASIT is a generally accepted method of treating IgE-mediated allergic rhinitis and asthma in patients over 5 years; the use of this method at an early age in the conditions of emerging immunity and tendency to Th2 responses is not studied enough yet.
2. Accumulating evidence indicates a relatively high clinical efficacy and safety of subcutaneous and sublingual immunotherapy in AR and BA in pediatric practice, however, there remains the necessity for additional research to confirm the effectiveness and long-term clinical benefits of using ASIT in children, especially at an early age.
3. There is evidence of ASIT preventive effect in children: the possibility to prevent the debut of BA in patients with AR, caused by tree and grass pollen, at least for the first two years after the end of ASIT.

ASIT is widely used in children, though the efficacy of this method in the conditions of the developing immunity and tendency to Th2 responses is being discussed.

Clinical recommendations and consensus documents define the age of 5 years and older as optimal for the possible start of ASIT. This is due,

first of all, to the characteristics of maturing an immune response in young children. It is known that the maturation of the immune system is most active during the early years and directly related to the effects of many infectious and non-infectious factors on the baby's body.

Peculiarities of implementing innate immunity mechanisms and a low functional activity of local immunological reactions in the first years of a child's life might cause not only high susceptibility to infectious diseases, but also imperfection of processes of tolerance formation [13]. The most important features of the immune response in young children are associated with functional immaturity of T cell component, prevalence of Th2 responses and low production of Th1 and Treg cytokines (IL-10, TGF- β), insufficient ability to differentiate B lymphocytes, effective antibody response (in particular, production of IgG and IgA) and the formation of immunological memory.

These features are exacerbated in children with atopic diseases due to further immune polarization towards Th2 response [14].

Nevertheless, a relatively high clinical efficacy and safety of subcutaneous immunotherapy (SCIT) and sublingual immunotherapy (SLIT) in AR and BA in children indirectly indicate the identity of immune response mechanisms to ASIT in adults and children. Moreover, only patients under 18 show preventive effect of SCIT and SLIT on BA debut in patients with allergic rhinitis [15].

ALLERGEN STANDARDIZATION

1. There are no common rules of standardizing allergens. There is no correlation and conversion formulas between different units of allergenicity.
2. Allergens, standardized in the biological way, are preferable.
3. The standardized drug must contain the basic major allergen proteins, however, such requirement in Europe should apply only to allergens of birch and timothy grass.
4. Standardized drugs of one manufacturer have a stable composition and reproducibility between series.
5. The result of clinical studies of therapeutic allergen can be fully applied only to a particular drug.

Allergen standardization is the expression of its allergenicity in common units in order to estimate its activity, risk and intended benefit of using as well as to prevent the emergence of drugs of inadequate quality. In contrast to chemotherapy drugs, it is impossible to standardize biological extracts only by their mass as the therapeutic allergen activity strongly depends on usefulness of raw extract and the degree of contamination of other proteins. The control of protein mass does not guarantee the preservation of allergen epitopes and drug immunogenicity [16]. Stages of allergen production are regulated by articles of the State Pharmacopoeia of the Russian Federation. It is easier to get a stable extract from pollen than from epidermal and fungal allergens. Mites require industrial cultivation as making extract from house dust leads to substrate contamination by various impurities [17]. Extracts from animal hair do not contain the whole range of protein molecules that cause symptoms, for instance, it is difficult to reach a stable concentration of prostatic dog arginase in the allergen of its epidermis [18].

Manufactures and regulators could not come to an agreement about using a unified methodology of standardization. In 2001–2008 during the CREATE project there was the study of 9 major allergens (Bet v 1, Phl p 1, Phl p 5a и Phl p 5b, Ole e 1, Der p 1, Der p 2, Der f 1, Der f 2) by which standardization of therapeutic drugs was expected. Nowadays the European Pharmacopoeia regulates the contents of only Bet v 1 and Phl p 5 [19]. The concentration of other major allergens in drugs is controlled by the manufacturer's internal quality standard. Content and activity of allergens in drugs must be in the range of 50–150% of the declared one.

The USA requires to express activity of all drugs in local units. Erythema d = 50 mm after intradermal administration of 0.05 ml of the solution is taken as reference D50 (or 100 000 BAU/ml). 19 allergen extracts have been created, which therapeutic allergens are compared with.

All the designations on medical allergens in the Russian Federation are based on the internal standards of their manufacturers. The vast majority of drugs pass biological tests after laboratory standardization. Stallergenes Greer for 100 IR (IR — standardization

unit, reactivity index) take a positive skin test $d = 7$ mm. ALK use standardized quality units SQ-U, based on data on the efficacy of the drug in clinical studies [20]. There is no direct comparison of drugs for ASIT as well as “coversine rates”. The instructions do not specify the number of major allergens, despite standardization on the content of essential proteins in the production. A domestic producer applies standardization on PNU without the use of biological techniques that may lead to differences in the activity of the drug in each series [21].

Nowadays questions of allergen standardization are not fully solved. It is not clear what dose of the allergen leads to an optimal development of tolerance as standardization units of different manufacturers are not comparable. Standardization on major components should solve some of the problems, however, the protein weight does not indicate its preserved structure and biological effects. The key indicator of the drug suitability for ASIT is sufficient clinical trials on a specific drug in the certain dosage according to the standardization, applied by its manufacturer.

REVIEW AND CLASSIFICATION OF THERAPEUTIC ALLERGENS

Drugs with registration in Russia are recommended for use, and strict adherence to the drug instructions is required.

Drugs, available in Russia, are made from natural raw materials and produced for parenteral (subcutaneous) and sublingual use (see Table 1).

In accordance with the legislation, the use of drugs, not registered in the regulatory documents of the Ministry of Health of the Russian Federation, is prohibited.

Therapeutic allergens of mold and epidermis of animals, insect poisons are not registered in Russia.

Allergens are drugs, obtained by extraction and purification of natural raw materials.

Allergoids are drugs from chemically modified protein molecules of the allergen. Allergoids cause fewer adverse reactions which is especially important in parenteral administration, however, there may be severe systemic reactions on allergoids. Allergoids are impossible to use for skin tests with a diagnostic purpose.

Injectable drugs for SCIT are presented in the form of water-salt solutions of allergens, allergoids and allergens, repository on the dosing vehicle. Repository provides a slower allergen release from the injection site, which increases safety; nevertheless, there may be anaphylactic reactions after applying similar allergens. Water-salt extracts are used on pre-season scheme, they are prescribed a few months before bloom and canceled not less than 2 weeks before the flowering period of cause significant plant. The indisputable advantage of repository forms is the possibility of their use on a year-round scheme (reducing the dose during the bloom of causal plant) and administration at the stage of maintenance of doses at intervals once in 4–6 weeks that simplifies conducting therapy and has almost no effect on the patient's social activity.

Table 1. **Drugs for ASIT registered in Russia***
Таблица 1. **Препараты для АСИТ, зарегистрированные в России****

Group of allergens	Name of drug	Characteristics
Dust mites <i>Dermatophagoides</i>	Dust allergen for diagnosis and treatment (FSUC “SIC “Microgen”)	SCIT
	Mite allergen <i>Dermatophagoides pteronyssinus</i> for diagnosis and treatment (FSUC “SIC “Microgen”)	SCIT
	Acarizax® (ALK-Abello A/C)	SLIT (<i>D. pteronyssinus</i> et <i>farinae</i>)
	LAIS Dermatophagoides (Lofarma S. p. A.)	Allergoid, SLIT (<i>D. pteronyssinus</i> et <i>farinae</i>)
	Staloral “Mite allergen” (Stallergenes Greer)	SLIT (<i>D. pteronyssinus</i> et <i>farinae</i>)
	Alustal “Mite allergen” (Stallergenes Greer)	SCIT, repository (<i>D. pteronyssinus</i> et <i>farinae</i>)
	Allergoid from house dust for treatment (FSUC “SIC “Microgen”)	Allergoid

Table 1. **Drugs for ASIT registered in Russia***
 Таблица 1. **Препараты для АСИТ, зарегистрированные в России****

Birch and trees of order Fagales	Staloral "Allergen of birch pollen" (Stallergenes Greer)	SLIT
	Fostal "Allergen of tree pollen" (Stallergenes Greer)	SCIT, repository, alder, birch, hornbeam, hazel
	Itulazax® (ALK-Abello A/C)	SLIT
	Mixed allergen from tree pollen for diagnostics and treatment (FSUC "SIC "Microgen")	SCIT
	Mixed allergoid of pollen alder, birch, hazel for treatment (FSUC "SIC "Microgen")	SCIT, allergoid
European ash (Fraxinus excelsior)	Allergen from European ash pollen for diagnostics and treatment (FSUC "SIC "Microgen")	SCIT
(meadow) grasses	Allergen from pollen for diagnostics and treatment (each as a monotherapy): <ul style="list-style-type: none"> timothy meadow herd grass (<i>Agrostis alba</i>) meadow brome (<i>Bromus erectus</i>) bluegrass (<i>Poa pratensis</i>) flint corn cocksfoot cereal ruttishness English ryegrass common foxtail (FSUC "SIC "Microgen") 	SCIT
	Grazax (ALK-Abello A/C)	SLIT, lyophilisate, timothy hay
	LAIS Grass (Lofarma S. p. A.)	Allergoid, SLIT, sift grass, timothy hay, bluegrass
	Oralair (Stallergenes Greer)	SLIT, gramen, spikelet, cockle, bluegrass, timothy hay
	Alustal "Allergen of meadow grass pollen" (Stallergenes Greer)	SCIT, repository, gramen, spikelet, cockle, bluegrass, timothy hay
	Pollen allergoid (each as a monotherapy): <ul style="list-style-type: none"> timothy grass cocksfoot meadow fescue grass (FSUC "SIC "Microgen") 	Allergoid, SCIT
	Mixed allergoid of pollen gramen, fescue grass and timothy hay for treatment (FSUC "SIC "Microgen")	Allergoid, SCIT
Weeds (different families)	Allergen from pollen for diagnostics and treatment (each as a monotherapy): <ul style="list-style-type: none"> ragweed absinth sage milk-witch gowan common sunflower (FSUC "SIC "Microgen") 	SCIT
	Pollen allergoid (each as a monotherapy): <ol style="list-style-type: none"> absinth sage ragweed (FSUC "SIC "Microgen") 	Allergoid, SCIT
	Ragwizax® (ALK-Abello A/C)	SLIT, lyophilisate, ambrosia
	Mixed allergen from pollen of weeds and sunflower for diagnostics and treatment (FSUC "SIC "Microgen")	SCIT, ambrosia, quinoa, wormwood, sunflower

* <https://grls.rosminzdrav.ru>, access time 08.06.2023

** <https://grls.rosminzdrav.ru>, время доступа 08.06.2023

In Russia drugs for SLIT are presented as a solution for sublingual taking, pills with the allergen/allergoid, absorbed on microcrystalline cellulose and lyophilized rapidly dissolving

pills. All these dosage forms provide an effective release of the allergen in the oral cavity where it penetrates through the mucous membranes and is captured by macrophages. Drops can be flexibly

dosed in the tolerability, however, they require compliance with the storage conditions and taking is associated with low compliance and failure probability, spontaneous dose change. Pills are more convenient, but indivisible. In case of reactions after taking the allergen, the doctor needs to adjust therapy, constantly assessing the balance of “risks and benefits”. The absence of certain “starting” doses with a reduced allergen concentration might increase risks of complications in highly sensitized patients. The advantage of lyophilizates over ordinary pills lies in their faster release [22].

Considering the provisions of EAACI consensus document, when conducting ASIT with pollen, it is possible to use the one of the “main” relevant plant in cases of patient sensitization to many plants of the same family, understanding, pollen will consist of homologous allergens [23]. As a rule, the main plant is most common, produces a large amount of pollen, contains main proteins of its group and causes symptoms in most patients [11, 24].

NEW AND PROMISING APPROACHES TO CONDUCTING ASIT

Approaches to ASIT optimization lie in changing route of allergen administration or changing allergen molecules for a safer and more effective application. It is proposed to use purer and more standardized extracts, recombinant allergens, allergen isoforms, oligopeptides of allergens, chimeras, fusion proteins, certain DNA vaccines, combinations with biological drugs, combinations with viral vaccines [25].

Alternatives routes of allergen administration: intralymphatic, epidermal, intranasal — aimed at improving the convenience of administration or optimization of the course (6–10 injections in intralymphatic route). Despite encouraging data

on clinical efficacy, research of their lasting effect is needed [26].

There is promising vector technology of allergen carriage by viruses and the combination of major protein molecules with lipoproteins, boosting immune response. There is development of vaccines, consisting entirely of major recombinant allergens that allows to reach exceptional efficiency and stability of the drug, particularly in the case of difficulty in producing and purifying natural extracts [27].

In the Russian Federation the group of Academician A. V. Karaulov develops a recombinant vaccine to treat cat allergy. It is shown that Fel d 1 cannot adequately bind IgE serums, therefore, there is study of efficacy of recombinant vaccine, containing uteroglobin and lipocalin for a wider coverage of polysensitized patients [28].

Clinical practice uses ASIT in combination with monoclonal drugs to achieve control in patients, who have not got ASIT due to the severity of the condition, and to modify the immune response, when used together. In the first case, any biological drug might be used, however, issues of ASIT efficiency remain open. Data were obtained for dupilumab on the possibility of its combining with ASIT in AtD [29]. Studies of preliminary omalizumab administration at high risk of ASIT anaphylaxis to food allergens have given positive results in the allergen tolerance, possibility to increase its starting dose and to reduce the risk of adverse events (AE) [30].

Insufficient randomized trials have been conducted to say that new allergens or routes of administration have a significant advantage over classical subcutaneous and sublingual ones. ASIT, combined with monoclonal antibodies must have clinical perspectives. Current drugs of monoclonal antibodies do not contain similar indications for the use in the instruction.

ASIT SAFETY, ADVERSE EFFECTS, WAYS TO IMPROVE THE SAFETY

1. ASIT when conducted by a specialist-allergist in accordance with the instruction for the use is a safe method of allergy therapy.
2. The incidence of anaphylactic reactions in ASIT depends on the type of allergen, route and rate of its administration as well as the condition of the patient's body.
3. Severe reactions to the allergen might be delayed, therefore, patients need to be monitored after SCIT up to 60 minutes.
4. Sublingual ASIT is safer than subcutaneous one. Cases of anaphylaxis in ASIT are of a casuistic nature.
5. Patients with systemic responses to SCIT can be successfully transferred to sublingual protocol.

The risk for severe reactions is considered one of the main ASIT drawbacks, especially exacerbations of asthma and anaphylaxis [31].

Adverse events of ASIT can be local and systemic. Local reactions are common both in SCIT (erythema, itching, induration and swelling at the injection site) and SLIT (oropharyngeal itching and (or) edema).

Systemic AE are more commonly associated with SCIT. Systemic adverse events occur less frequently in children than in adults.

AE, requiring discontinuation of therapy or significant dose adjustment, drug change or route of administration, include: anaphylaxis, reactions, requiring the use of epinephrine, severe edema of pharynx and oral cavity, eosinophilic esophagitis, severe exacerbation of BA.

Particular attention should be paid to the fact that 72.4% of all systemic responses are delayed and occur 30 and more minutes later after the allergen administration [32]. It is necessary to review guidelines in the direction of extending the follow-up period after the allergen administration and a mandatory adequate patient instruction.

A favourable safety profile of ASIT is due to the peculiarities of sublingual capture and allergen processing. Allergens penetrate through the mucous membrane, where are captured by

tolerogenic antigen-presenting cells, after which there is allergen introduction for recognizing by T lymphocytes [33]. In the absence of damage and inflammatory process, systemic penetration of the allergen and its recognition by mast cell receptors are less probable in the tissues of the oral cavity [33]. Subcutaneous administration route is associated with a greater risk of the allergen contact with circulating pro-inflammatory basophils and Th2 lymphocytes [32–34].

Patients, experiencing serious adverse reactions in SCIT, could potentially be transferred to SLIT [33].

Of particular interest are factors that increase risks of systemic responses and anaphylaxis, first of all, the dose of allergen and dose regimen. A dose-dependent effect is lower in SLIT, but there is a faster dosage adjustment in SCIT than it is recommended by the manufacturer; erroneous administration of increased dose of the allergen is associated with an increase in the frequency of anaphylaxis [11].

S2k guideline for ASIT, accepted by professional allergy associations of Germany, Austria and Switzerland, highlights the factors of developing systematic responses [35]:

1. allergy symptoms at the time of treatment, possible allergen exposure;
2. current infection;
3. mastocytosis; hyperthyroidism;
4. high sensitization rate;
5. inadequate increase in the dose of the drug in the initial phase of treatment; an allergen overdose;
6. administration of certain drugs (beta-blockers, angiotensin-converting enzyme inhibitors (ACE));
7. inadequate circulation load: excessive alcohol consumption, excessive physical exertion, visiting the sauna;
8. violation of the injection technique;
9. non-compliance with the manufacturer recommendations for reducing the dose when transferring to a new batch.

If there are indications, the recommended regimen of SCIT might be changed into a more sparing and cautious [36].

Oropharyngeal injections and lesions (ulcer, gingivitis, stomatitis) might be potential risk factors in SLIT due to possible systematic allergen penetration; on the other hand, it is not clear what effect immune responses have on the course of recurrent stomatitis during the allergen administration. Inflammatory diseases of the oral cavity contraindication for SLIT [11, 33, 35].

Risk patterns of adverse events in ASIT can be traced in connection with the type of therapeutic allergen used. The risk of systemic responses in ASIT is lower in patients with sensitization to dust mites (DM) and higher in pollen polysensitization (more than 3 allergens), higher for extracts of cereal grass pollen, compared to allergoids [31]. Extracts, containing stable proteins – “anaphylaxis molecules”, and sensitization to minor proteins (usually coinciding with polysensitization) are associated with a higher risk of anaphylaxis. Such severe reactions are characteristic of immunotherapy with extracts of weeds, nuts and cereals.

AE treatment is carried out on the general principles, according to nosological form. Local AE can disappear on their own or with the use of antihistamines through the mouth. Non-life threatening angioedema can be relieved using systemic steroids once. Symptoms of BA (not as part of anaphylaxis) are relieved with the combination of inhaled bronchodilator and corticosteroid. In the presence of anaphylaxis criteria there is administration of epinephrine and treatment on algorithms of anaphylaxis cupping.

In Russia ASIT is carried out only by doctors allergist-immunologist, experienced in this type of therapy and capable to provide emergency relief with the development of an allergic reaction.

Allergist's offices should be equipped to provide necessary emergency assistance, and patients should be fully informed about the possible risk

of adverse reactions, that should be documented. It is unacceptable to delay with epinephrine administration in the development of anaphylaxis [11]. Written informed consent for treatment must be obtained from patients, similar to the one, used for vaccination.

It is recommended to increase follow-up time up to 60 minutes after the allergen administration.

The use of drugs, which have passed biological standardization, safety and efficacy of which are confirmed in clinical studies, allows a relative confidence in the stability of the composition and activity of the allergen in the drug [11, 37]. It is necessary to evaluate the patient's condition before each injection [38].

CONTRAINDICATIONS TO ASIT

1. Conducting ASIT to the patient with relative contraindications justified if the expected benefit of treatment exceeds the possible risk of deterioration.
2. Prior to ASIT there should be spirometry and evaluation of dynamic peakflowmetry results in persons with BA throughout the treatment period.
3. The presence of autoimmune diseases is a relative contraindication to ASIT during remission and an absolute contraindication in the active stage.

Contraindications to ASIT can be absolute and relative. Conducting ASIT to the patient with a relative contraindication is justified if the expected benefit of treatment exceeds the possible risk of patient deterioration [37, 39, 40].

In 2015 EAACI published a position paper, presenting contraindications to ASIT [39]. There was no later revision of this document.

ASIT is not used under 2 years. In children from 2 to 5 years a decision can be taken on an individual basis, based on the child's quality of life and adherence to therapy. However, there are no drugs of therapeutic allergens in Russia, allowed for use in children under 5 years. No other age group is in itself a contraindication to ASIT [11, 39–41].

Absolute contraindications to ASIT [11, 37, 39, 40, 42–44]:

1. Severe or uncontrolled BA, forced expiratory volume in 1 second (FEV₁) is < 80% in children [11],
2. malignant neoplasms in the active stage,
3. autoimmune diseases in the active stage,
4. initiation of treatment during pregnancy,
5. poor compliance with treatment.

Relative contraindications to ASIT:

1. Partially controlled BA.

Patients, experiencing severe or medically uncontrolled asthma, are at an increased risk of systemic responses by aeroallergens in ASIT (mainly in SCIT) [45]. Spirometry is necessary before starting ASIT. Before each injection patients with BA should be assessed for the disease control rate measured for peak expiratory flow. In the event that lung function has decreased more than 20% of the best individual value, the injection should be postponed even if these indicators meet age and race standards [36, 39]. There is no evidence that ASIT can exacerbate asthma severity or induce it de novo.

2. Immunodeficiency, HIV infection, taking immunosuppressants, anticancer agents, chronic infections.

Each immunodeficiency has individual pathological mechanism, and in addressing the issue whether the patient should be treated with ASIT, the key feature is evaluating potential efficacy [39].

Some guidelines indicate concomitant treatment of patients with immunosuppressants as the contraindication to ASIT since these drugs might have a significant negative impact on the effectiveness of therapy.

HIV infection is a relative contraindication to ASIT. HIV infected patients, receiving antiretroviral therapy, can take ASIT in the early stages of the disease with the level of CD4 > 400 cells/mm and an undetectable viral load [39, 46]. Any stage C disease (according to CDC-classification of 1993) is considered absolute contraindications to ASIT [46].

Chronic viral infections (hepatitis B or C in remission) as contraindications to ASIT are not listed in the literature [39].

ASIT should be prescribed depending on patients' individual characteristics, considering states of immunodeficiency or the course of chronic infection as relative contraindications.

3. Psychiatric/mental disorder, impairing cooperation between doctors and patients.

SLIT may be considered in a child with a mental disorder only if it is conducted with a controlling guardian [11, 39].

4. Autoimmune diseases in remission or organ-specific autoimmune diseases.

Some guidelines consider the presence of autoimmune diseases a relative contraindication to ASIT, others — absolute. Controlled trials haven't identified an increased risk of autoimmune disease manifestation amid ASIT in patients with allergy. In case of developing an autoimmune disease, ASIT should be discontinued, and it shouldn't be prescribed to patients with autoimmune diseases in the active stage [39]. Hashimoto compensated thyroiditis against the background of drug therapy is not a contraindication to ASIT. In multiple sclerosis, myasthenia gravis, systemic lupus erythematosus, rheumatoid arthritis, Crohn's disease the decision on ASIT can be made

on an individual basis in view of no disease activity [11, 40, 43, 47].

5. Chronic diseases.

Some concomitant diseases raise concern when considering the possibility of ASIT due to the lack of any specific information.

Cardiovascular disease in an unstable or progressive stage is a relative contraindication to ASIT with inhalation allergens. Before starting ASIT, there should be a careful evaluation (preferably with a cardiologist) of cardiovascular disease status, its therapy and the risk of anaphylaxis (requiring epinephrine use) [11, 39].

6. Malignant neoplasms.

Malignant neoplasms are considered absolute contraindications to ASIT. Long-term remission and the medical consultation with the oncologist allow to consider ASIT with strict indications and assessment of all risks.

7. Use of drugs.

Drugs of β -blockers, angiotensin-converting enzyme inhibitors and monoamine oxidase can change the effect of epinephrine in the development of anaphylaxis. The use of β -blockers in pediatric practice is limited to rare cases in rhythm disturbances and ophthalmologic pathology. If possible, β -blockers should be replaced by alternative drugs. If replacement is not possible, ASIT is allowed with careful assessment of risks and an individual dosage regimen [38, 40].

8. Severe form of atopic dermatitis.

In European guidelines ASIT might be considered as a potential treatment in patients

with severe AtD, associated with other allergic diseases with sensitization to HDM, birch pollen, meadow grasses as well as epidermal allergens if contact with the allergen cannot be avoided [43, 48].

9. Eosinophilic esophagitis (EoE), chronic inflammatory gastrointestinal diseases.

According to statistics, EoE may occur in patients, with pollen allergy with the manifestation of oral allergy syndrome. SLIT is contraindicated in patients with diagnosed EoE [49]. There is increased risk of EoE in patients, receiving SLIT due to ingestion of the allergen and local contact with immune cells of the esophageal mucosa. SCIT prescription is possible in EoE remission [49]. SLIT is not used in patients with chronic inflammatory oral diseases [43].

10. Severe systemic responses to ASIT in anamnesis.

Documented episodes of anaphylaxis during SCIT are its contraindication [33, 43, 50]. Given higher safety of SLIT, there is an opinion that patients with severe adverse responses to SCIT can be shifted to SLIT [33].

11. Pregnancy.

ASIT initiation during pregnancy is contraindicated. If before pregnancy the patient has received and tolerated ASIT well, this type of therapy may be continued during childbearing (following all precautions) [39].

As temporary contraindications to ASIT, most guidelines consider acute infections of the respiratory and gastrointestinal tract, tooth extraction, oral surgery, exacerbation of allergic diseases [39, 40, 37, 43].

EFFICACY OF ALLERGEN-SPECIFIC IMMUNOTHERAPY

1. ASIT efficacy is proven in diseases, causally associated with sensitization to HDM and plant pollen.
2. There is plenty of evidence of ASIT effectiveness in AR, BA.
3. ASIT is not included to the standard of therapy in AtD, however, with a combination of AtD and allergic rhinitis therapy reduces symptoms of both diseases.
4. The minimum duration of therapy is 3 years. There is evidence of maintaining ASIT effect after the end of therapy.

ASIT WITH ALLERGY TO HOUSE DUST MITES

Indication for ASIT with HDM allergen is proven clinically relevant sensitization. The treatment is carried out according to the instructions, dosage depends on the method/form of therapeutic allergen release [51].

All HDM belong to the same group and have a similar structure. Particles of chitinous cover, faeces and eggs are sources of allergens. The predominant type is mites of the genus *Dermatophagoides* with 39 described allergens. Isolated sensitization to non-dermatoglyphic mites occurs only in 1.5% of cases, in others, it is combined with *Dermatophagoides*. Clinical efficacy and safety of ASIT is shown with the allergen of mites *Dermatophagoides* in children with BA and AR, in the presence of cosensibilization to mites *Blomia*, *Lepidoglyphus* regardless of the therapy method [52]. Apparently, this is due to the fact that despite standardization of therapeutic allergens on Derp (f) 1 and Derp (f) 2, they also contain other proteins, which may serve as a possible reason of efficiency in sensitization to other mite allergens.

ASIT with HDM allergen in BA is the only type of immunotherapy, included in the guidelines of Global initiative for Asthma, GINA. The Cochrane meta-analysis data show ASIT efficacy in children in terms of BA symptoms and a high safety profile of sublingual treatment method [53]. ASIT efficiency in children with AR is confirmed by a decrease in the severity of symptoms and reduced need for drug therapy [54].

Immunotherapy with HDM is an additional method of treatment in children with AtD and (or) combined with BA, AR it is recommended when there is a connection between exacerbations of the disease

and allergen exposure [55]. ASIT with HDM drugs improves the course of AtD at any age, especially in monosensitization [56, 57].

According to guidelines, the optimal duration of ASIT is not less than 3 years. The clinical effect of ASIT with HDM allergen, when used in sufficient doses, develops after the first 6 months of therapy. It is possible to trace maintenance of the effect during 5–7 years after the end of ASIT [58].

Therapeutic HDM allergens for SLIT are produced in the form of drops (Stallergenes Greer) and pills, containing the allergen (“ALK-Abello A/C” manufacture) or allergoid (“Lofarma S. p. A.” manufacture). Efficacy and safety of these drugs are shown in children of different ages with AR and AR, combined with BA [59–62].

ASIT with HDM allergen is effective against symptoms of BA, AR and AtD, caused by relevant allergens in children with mono- and polysensitization.

ASIT IN POLLEN ALLERGY

Pollen allergy is one of the most widespread types of hypersensitivity in children. The main manifestations of pollen allergy are AR (with conjunctivitis) and BA.

The Cochrane analysis by M. Abramson (2010) was among the first which showed SCIT advantage over placebo (27 studies) in pollen allergy [63]. The meta-analysis by Fortescue R. (2020), studying the effect of SLIT on BA, did not provide the division by the allergen type, however, SLIT had an advantage over placebo in the vast majority of studies [53].

Long-term practice as well as domestic studies have proven that SCIT with pollen allergoids and allergens of the Russian production is effective method of treating children with AR (rhinoconjunctivitis) and BA [64]. Pre-season course is used (the beginning of administration 4 months before flowering) in treatment with subcutaneous drugs of water-salt pollen allergens and allergoids. Allergen mixes are used with allergy to several allergens of the same group. Nevertheless, the dose of each allergen in the mixture is reduced that makes it difficult to reach the optimal dose of the main allergen. The initial course of treatment includes 32 injections for an allergen and 25 for an allergoid. The rhythm of administration is chosen depending on the drug tolerability; the dose is not increased in case of local reactions, the

reduced dose is repeated until satisfactory tolerance is achieved. After reaching maximum dose, injections are repeated with at intervals of 5–7 days prior to the beginning of the pollen season. Accelerated schedules of SCIT with domestic allergens (“fulminant” schedule) is not recommended by APAIR experts for safety reasons.

Deposited allergens “Phostal” — the extract from the mixture of tree pollen and “Alustal” — the allergen of meadow grasses (Stallergenes Greer) are not available for use at the time of the article issue. After increasing a dose for about 4 months, a maintenance dose is administered all year round at intervals of 2–6 weeks during 3–5-year course of therapy; the dose is reduced half or more during the flowering season.

Pollen allergens for SLIT exist in two forms — drip and pill. Staloral “Allergen of birch pollen” is available in the form of drops, used for pre-season-seasonal protocol. The advantage of drops is the possibility of flexible dosing with reaching the maximum maintenance dose of 240 IR daily that is the key to optimal efficacy and implementation of the modern approach of high-dose SLIT. Efficiency of drops with the allergen of birch pollen is proven in a double-blind placebo-controlled trial, using a daily dose of 300 IR in pre-season-seasonal therapy mode (574 patients) [65]. Similar efficacy and safety of SLIT with the allergen of birch pollen is shown in patients with oral syndrome (cross food intolerance) as well as without it.

Sublingual drug of birch pollen in the form of lyophilized pills 12 SQ is registered in Russia for treating AR and (or) rhinoconjunctivitis in children 12 and older. Currently Itulazax drug is passing the final stage of the trial in the population of children aged 5 years in Europe.

In our country drugs in the form of pills are available for SLIT of allergy to cereal grasses: the pollen allergen of 5 herbs (“Oralair”), the allergen of timothy grass in the form of lyophilized pills (“Grazax”) and carbamylized allergoid from a mixture of 3 herbs (“LAIS Grass”). Treatment is carried out according to pre-season-seasonal protocol, “Grazax” 75000 SQ-T drug can be used all year round.

Current EAACI guidelines allow to use one allergen for treating diseases, caused by homologous

plants. For pollen of cereal herbs the major allergen is proteins of timothy grass Phl p 1 and Phl 5 b. The study of antibody neutralizing capacity shows that allergens of major cereal herbs are homologous and bind in vitro by 98–100% [66]. It allowed to use extracts from the allergen of timothy grass for effective immunotherapy for patients with allergy to cereal grains.

In pediatric practice GAP study (n = 712) has shown high efficacy and safety of “Grazax”, similar to that in adults, as well as ASIT effect with the pollen of timothy grass on the development and course of allergic asthma. Significant reduction in symptoms of existing asthma, amount of therapy received and improvement in FEV₁ were noted. Effect from ASIT regarding asthma exacerbations, induced by grain pollen, lasted for minimum 2 years after the end of treatment [67].

Treatment protocol of the drug “Oralair”, containing 5 herbs (cocksfoot, vernal grass, common ryegrass, bluegrass, timothy grass) involves the use of high maintenance doses of 300 IR/daily. Efficacy and safety of the drug in pediatric practice are shown in a multicentre double-blind placebo-controlled trial (278 children and adolescents aged 5 to 17) with a decline of 28% in the index of nasal and ocular symptoms [66].

Monomeric allergoid of cereals 1000 AU, “LAIS Grass”, was studied in several small groups, which results are summarized in the meta-analysis by Mosges R. (2010). The average change in the scale of symptoms and need for medication was compared with placebo and amounted to –34% and –49%, respectively. Serious adverse reactions were not reported, all AE were of a local nature [62].

The standardized drug “Ragvizax” is available in Russia in the form of lyophilized pills 12 SQ with the allergen of ambrosia, registered for patients 5 years. In pediatric practice the drug was studied in the population of 1002 children aged 5–17, having clinical manifestations of AR or AR, combined with BA; there was a decrease of 47.7% in the amount of drug therapy [68].

In clinical trials polysensitized patients noted good tolerability and the possibility to combine the drugs of timothy grass 75000 SQ and ambrosia 12 SQ when taking pills with the difference in 5 minutes [69].

ASIT is the only pathogenetically substantiated method of treating respiratory allergic diseases, associated with sensitization to plant pollen. ASIT gives the patient a chance to prevent the aggravation of the process, expansion of the spectrum of sensitization as well as a chance to achieve a stable remission.

METHOD OF ASIT

The key steps in conducting allergen-specific immunotherapy:

1. Verification of the diagnosis and identification of evidence for ASIT.
2. Identification of a clinically significant allergen via allergological examination.
3. Determination of contraindications to ASIT.
4. Drug selection of therapeutic allergens.
5. Initiation of ASIT during remission of allergic diseases; the first administration of allergy vaccine in the allergist-immunologist's office.
6. Achieving maximum compliance with parents, training the technique of taking the allergen and self-help with the development of adverse reactions, drawing up an individual action plan.
7. Monitoring the efficiency of therapy.

ASIT is indicated in a positive skin test or in identifying an increase in the level of specific IgE (it is believed that sensitization must at least comply with the moderate class) to a suspected allergen. In some cases of polysensitization it is necessary to determine major components of plant pollen to choose an allergen for treatment.

Before ASIT it is necessary to revise the diagnosis, to eliminate the effect of comorbid conditions, occurring with similar symptoms, and to ensure that complaints and symptoms, which disturb the patient, are associated with sensitization.

Remission of the underlying disease is required to start a course of ASIT. It may be spontaneous (for instance, in winter for pollen allergy) or achieved on the basic therapy.

In BA the course of the disease should be monitored for minimum 1 month (it is the minimum period for evaluation of asthma control). By the start of ASIT the patient with asthma should also have acceptable lung function rates, in particular, FEV₁ in children must be at least 80%

of the predicted. Low FEV₁ is not only the rate of uncompensated asthma, but also the risk indicator of possible systemic responses to ASIT [11, 70].

SLIT is possible with the oral mucosa integrity and lack of infection foci, therefore, it is crucial to inspect the oral cavity at each visit and train parents to detect trauma, ulcers and erosions. In the event of defects in the mucous membrane, SLIT should be suspended. It is necessary to determine the possibility of having SLIT against the background of orthodontic intervention, implantation, bracket system for the correction of the malocclusion in each specific case.

Before treatment it is required to reveal the patient and their parents the principle of ASIT action, goals of treatment (especially long-term points of efficacy), differences from pharmacotherapy of allergic diseases, peculiarities of treatment regimen, forthcoming labor, time and treatment costs, duration, measure of family responsibility for implementing treatment scheme, precautionary measure, possible AE. Information may be provided both orally and in the form of brochures. The first visit requires signing the informed consent form, training the technique of the drug administration, providing the patient with a written self-help plan in developing AE. The diet should be complete, age-appropriate, but in case of significant food allergy and anaphylaxis in the anamnesis, it is necessary to ensure elimination of the allergen. When conducting ASIT in the flowering season, it is advisable to implement the recommendations on maintaining hypoallergenic household and minimizing contact with pollen. During SLIT it is reasonable not to use food with irritating effects and traumatic for mucous tunic of the mouth.

It is advisable to perform vaccination against infectious disease, according to the vaccination schedule, 1 month and more before the start of ASIT.

The first administration of the allergy vaccine to the patient, regardless of the route and dosage form, is conducted in the allergist-immunologist's office. This also applies to repeated courses, carried out by pre-season-seasonal protocol and to year-round courses, reinitiated after a long break. Before the

first administration, the patient's admission to the beginning of treatment is registered in the patient's medical record: the absence of contraindications, examination data, the lack of data on intercurrent communicable disease. SLIT requires data on the inspection of the mucous membrane of the mouth, the absence of inflammation, lesions. After the first administration of the allergy vaccine, the patient should be monitored at least 30, preferably 60 minutes.

SCIT at home is prohibited both in the dosage phase and conducting maintenance therapy!

Patients, receiving SCIT, follow the injection schedule, indicated in the instructions for the drug. Patients, receiving SLIT, require the follow-up schedule. The critical period of SLIT is the first month of therapy. At this time, as a rule, patients note the emergence of local AE and, due to having no way to contact the attending physician, many parents discontinue treatment because of "intolerable adverse events". Thus, the follow-up of the patient, who has started the course of SLIT, should include:

- availability of the doctor or a competent nurse for the patient, for example, by phone, for quick resolution of emerging issues;
- preliminary explanatory work on the course of treatment before its start;
- follow-up visit during the first month of therapy to assess tolerability of treatment; then — visits to assess the course of treatment, compliance, tolerability of treatment, discussion of the vaccination schedule and etc. — each 3–6 months;
- there may be off-plan visits in case of intercurrent diseases, exacerbations of the underlying disease and questions from patients;
- regular evaluation of ASIT treatment efficacy with HDM is made once a year, with pollen allergens — after the flowering season of plants casual;
- decisions, made on visits: about the possibility of reducing the amount of pharmacotherapy of the underlying disease as well as the moment of discontinuing ASIT [11]. AE are monitored at each patient's visit.

Temporary interruption of ASIT course occurs for various reasons:

- acute intercurrent disease;
- exacerbation of the underlying disease; in patients with BA — decrease in peak expiratory flow up to 80% of the maximum and lower ("yellow zone");
- organizational reasons (departure, untimely purchase of another package of allergy vaccine, etc.);
- in ASIT: a violation of the integrity of the mucous membrane (stomatitis, aphtha, injury, tooth loss/extraction); teething is not a contraindication to continue SLIT if it is not accompanied with bleeding, inflammatory process; acute gastritis, gastroenteritis.

In classical schemes of SCIT with domestic water-salt allergens, in case of the patient's turnout after an approved break, the next dose should be reduced, that is, "to back down" under the scheme of ASIT by 2–4 doses. It is required to "to step back" by 1 dilution when interrupting the course for more than 14 days.

One should be guided by the instructions on resumption of therapy after interruption, given by the manufacturer, during SCIT and SLIT with standardized allergy vaccines. There is no dosage reduction during SLIT with tableted vaccines.

Vaccination against infectious disease in patients, receiving ASIT, should be conducted within periods, as close as possible to the vaccination schedule. Vaccination is not carried out in the stage of building up the dosage of an allergy vaccine (in ASIT with domestic water-salt allergens, parenteral allergens Phostal/Alustal). In treating with sublingual vaccines, which dose build-up stage is 3–9 days or completely absent, we do not recommend to conduct scheduled vaccinations against infectious diseases in the 1st month of SLIT.

In SLIT at the stage of maintenance therapy, vaccination requires a temporary interruption of taking allergenic drug: 3 days before the intended vaccination, on the day of vaccination and for 10–14 days after vaccination [71]. In case of an emergency vaccination or the one for epidemic indication, it is also necessary to check the instruction for the drug.

EVALUATION OF ASIT EFFICACY

1. Biological markers are molecules, cells, receptors, detected in the blood or other biological body fluids and being the measure of the severity of the disease and (or) its response to therapy.
2. Nowadays there are no biomarkers available in clinical practice, reflecting the onset of the therapy effect.
3. Medical practitioner regularly assesses ASIT efficacy, based on changes in symptoms and the amount of drug therapy.
4. There is no standard rating scale, accepted by all communities.

Since ASIT is not fully effective for each patient, the identification of patient's biological markers is of great importance to predict clinical efficacy of immunotherapy and monitor response to treatment.

Of particular importance in predicting ASIT efficacy is specific IgE to the major allergen [41]. Interrelations are described between the profile of molecular sensitization and response to ASIT. Thus, it is shown that sensitization to Der p 1 or Der p 2 might be a good predictor of SLIT efficacy with HDM allergen [72], and combined sensitization to Phl p 5 and Phl p 12 predicts the development of AE [73]. ASIT is not appropriate with no sensitization to the major component of a casually significant allergen.

In 2017 the working group of EAACI, "Biomarkers for monitoring clinical efficacy of allergenic immunotherapy" identified potential biomarkers [74]:

— IgE. There was a demonstrated increase in specific IgE during the first months of ASIT with its subsequent progressive decline after 6 months of therapy. At the same time, there are contradictory data on the significance of the ratio in levels of specific to total IgE as a predictor of clinical response to ASIT [10].

— IgG. Repeated exposure to the allergen induces the formation of allergen-specific IgG4 antibodies, blocking action of which is currently considered one of the main ASIT mechanisms for respiratory allergy. The correlation is shown between the production of specific IgG4 antibodies and clinical outcome of ASIT. Nevertheless, specific IgG4 cannot be currently be considered as a reliable biomarker of ASIT efficacy due to the need for long-term studies.

— Inhibitory activity of serum against IgE (IgE-FAB) is regarded as a potential biomarker of ASIT efficacy, but it is not used in practice.

— Basophil activation test has been suggested as a potential biomarker of ASIT efficacy, but the results of the work are contradictory [74, 75].

— There is active discussion of using chemokines, cytokines, cells, participating in an allergic response, for instance, producing IL-10 Treg lymphocytes, molecular markers of dendritic cells, however, they are difficult to use in practice.

— Provocative tests, undertaken in dynamics, are attributed to "Biomarkers in vivo". Thus, endonasal provocative test is used as an indicator of ASIT efficacy in its clinical studies.

Therefore, there is no current validated approach to the use of biological markers to confirm ASIT efficacy in clinical practice.

A the present stage the only possible way to assess ASIT efficacy is the application of clinical efficiency criteria: 1) the reduction in the severity of the disease symptoms, caused an allergen, with which ASIT is carries out, the reduction in the period of exacerbation; 2) the reduction in the need for medication; 3) change in the natural course of allergic disease (stable remission that persists after the end of therapy; prevention of expanding the range of sensitization; prevention of developing new clinical forms of atopic disease).

Satisfaction with treatment is assessed in patient with BA according to the scheme proposed by A. D. Ado [76], and for patients with AR — on a similar principle: 4 points — an excellent effect — disease remission; 3 points — a good effect — very rare and mild symptoms, a sharp decrease in the need for medication; 2 points — a satisfactory effect — exacerbation of the disease is less frequent, symptoms — milder; 1 point — no effect. Scoring is quick, simple and convenient.

Clinical studies of ASIT apply integrated index, addressing symptoms of allergic diseases and the necessary amount of pharmacotherapy, for instance, recommended EAACI scale to determine the average score of nasal and conjunctival symptom severity and the need for drug therapy to relieve symptoms (Table 2) [77].

EAACI clinical guidelines for ASIT in asthma of tick-borne etiology provide "The list of positive changes during AIT with HDM in tick-borne asthma" [42].

It is certainly important to register epicrisis in patients' medical records, where a practical physician reflects the integrated assessment of complaints, symptoms, absences in the child care institution and disability, basic and emergency therapy, antibiotic prescription for respiratory tract infections, an emergency visit to an ENT specialist (with the manipulation), hospitalization and etc.

Table 2. **Index calculation table CSMS (combined symptom and medication score)**
 Таблица 2. **Таблица подсчета индекса CSMS (combined symptom and medication score)**

Nasal	Itching in the nose	0–3	0 — No symptoms 1 — Mild, easily tolerated, do not affect the well-being 2 — a clear feeling of a symptom, worries constantly 3 — Intolerable, an impact on sleep, daily activity
	Sneezing	0–3	
	Discharge from the nose	0–3	
	Stuffiness	0–3	
Conjunctival	Itching/redness	0–3	3 — Intolerable, an impact on sleep, daily activity
	tearing	0–3	
Daily symptom score (DSS)*		0–3 (number of symptoms/N signs)	To assess symptoms on HDM eye symptoms are not considered
Assessment of DMS therapy	Antihistamine (locally or systematically)	1	
	Intranasal steroids	2	
	Oral steroids	3	
Overall assessment of daily therapy received (DMS)		0–3	
CSMS — combined assessment of symptoms and therapy	DSS (0–3) + DMS (0–3)	0–6	

The timing of the efficacy assessment are essential. In treating with HDM allergens mandatory cutoff value for initial efficacy assessment is 1 year from the start of therapy. In the absence of effect after the first year/course of ASIT it is necessary to check: the course dose of the allergen by counting PNU when treating with water-soluble allergens or the number of packages of sublingual drugs, patient compliance with a treatment regimen, a break in treatment; re-evaluate the diagnosis, indications for ASIT, the correctness of allergen choice for therapy; make a decision to continue or terminate ASIT [11].

ATOPIC DERMATITIS AND ASIT

1. Atopic dermatitis is not an obstacle to ASIT in patients with AR, BA.
2. In patients with isolated AtD in case of clinically significant sensitization and association of exacerbations with a specific inhaled allergen, ASIT must bring a clinical effect, yet, the indication of AtD is not included in the guidelines to drugs for ASIT, registered in Russia.

ASIT is indicated for patients with an identified cause-significant allergen, responsible for the development of disease exacerbations. In AtD it is often impossible to identify the main cause of exacerbations as xerosis, mechanical damage to epidermis, change in skin microbial combination as well as non-specific triggering play an important role

in the deterioration of the skin process [55]. Besides, most patients have non-IgE-mediated phenotype of AtD, in which ASIT will be ineffective.

In Russian clinical guidelines ASIT with pollen and household allergens is recommended to patients with allergic respiratory diseases and concomitant controlled AtD and patients with AtD with no respiratory allergy and proven cause-significant sensitization to HDM allergens [55, 78].

The consensus of the European Association of Dermatovenerologists for treating AtD points that in this pathology ASIT might be considered in patients with severe disease, sensitized to birch pollen, meadow grasses and HDM and the presence of exacerbations in the anamnesis, coinciding with exposure to an allergen [48].

Research findings on efficacy and safety of ASIT with aeroallergens in children with AtD were published in recent years. Systematic reviews and meta-analysis show good efficiency of ASIT with pollen and household allergens in patients with AtD in decreasing SCORAD index and the need for topical glucocorticosteroids [79–81]. There are encouraging trials of ASIT efficacy in adult patients with AtD, sensitized to allergens of cat and dog dandruff.

Thus, after achieving clinical remission, AtD cannot be contraindication for immunotherapy to treat concomitant AR and (or) BA. In patients with

isolated AtD when associating exacerbations with specific inhaled allergens, ASIT should have clinical effect, yet, AtD indication is not included in the instructions of drugs for ASIT, registered in Russia.

ASIT IN FOOD ALLERGIES

1. ASIT is recognized as an effective method to develop tolerance to the allergens of milk, eggs and peanuts in children over 5 years.
2. Standardized drugs of food allergens for treatment, methodologies of a food provocative test and methods of ASIT with food allergens are not presented in Russia.
3. Overall, work on conducting oral immunotherapy in food allergies are experimental.

The standard of food allergy/anaphylaxis treatment remains avoidance of food allergens, however, recently there has been an increase in ASIT studies both to prevent food allergies and to consider as a method of treatment. The first double-blind placebo-controlled trial of SLIT with food allergies was published in 2005. During therapy 45% of patients in the treatment group reached the highest dose of hazelnut (20 g) as compared with 9% in the placebo group [82].

EAACI recommends oral immunotherapy (OIT) as a promising therapeutic method with high short-term efficacy regarding allergy to milk, eggs and peanuts since the age of 4–5 years, however, does not provide recommended schemes of food introduction or the use of standardized drug for OIT. Pharmaceutical companies do not produce therapeutic drugs of food allergens, making it difficult to widespread the method [83, 84].

Anaphylaxis and EoE are referred to possible risks of food ASIT; there is no full information on the duration of maintaining tolerance to food allergen, in other words, on the risk of food allergy relapse after terminating regular intake of a maintenance dose of food allergen. Before wide implementation of food ASIT in practice it is required to find and understand markers, which differentiate desensitization and sustained resistance, and to introduce them for monitoring immunotherapy.

It is necessary to standardize the most important food allergens (anaphylaxis molecules ovomucoid Gal d 1, casein Bos d 8, peanut Ara h 2 и Ara h 6, hazelnut Cor a 9 and Cor a 14, cod parvalbumin Gad c1, shrimp Pen m 1, Pen m 2), with which treatment is justified when symptoms are severe (food anaphylaxis) and the cause is hard to avoid.

With OIT (compared to other forms of immunotherapy) quite large total doses of allergen are used, and thanks to OIT patients can get protection not only from dangerous responses due to accidental exposure of trace amount of allergen, but also from reactions after consuming gram amounts of allergenic products.

In 2020 Food and Drug Administration US (FDA) approved a standardized product for OIT (Palforzia™) to treat food allergy to peanut and also reported on programs for treating allergy on eggs and walnut [85].

Desensitization immunotherapy with food allergens currently includes: oral, sublingual and epicutaneous immunotherapy, using native food allergens or recombinant proteins affected by mutagenesis or over-heated food. Intranasal, intralymphatic and epicutaneous allergen administration is referred to experimental methods of ASIT with food allergy.

The main problem is the lack of consensus regarding doses of the allergen, schemes of its administration, oral provocative testing methods in children as well as failure of reached tolerance after a break/discontinuation of OIT. There is study of conduction OIT combined with anti-IgE monoclonal antibodies to improve safety and tolerability of ASIT. To date, omalizumab is not registered to treat anaphylaxis and food allergy. The earliest age to use is defined as 6 years for allergic asthma.

USE OF SYMPTOMATIC DRUGS AND ASIT

Antihistamines, due to effect on mast cells and reaction of immediate type, as well as glucocorticosteroids, inhibiting the late phase of inflammation, could potentially interfere with efficacy of immunotherapy and incidence of AE.

According to national and international clinical guidelines, ASIT is recommended to children with AR during remission, including drug-induced one, and to patients with a mild and moderate BA, controlled with pharmacotherapy [11, 41]. Most protocols of ASIT studies assumed that during therapy the patient may get symptomatic drugs, if necessary, and under these circumstances immunotherapy appears effective. Inhaled glucocorticosteroids have minimum systemic effect, do not impact on efficacy of immunotherapy and the formation of humoral protection factors [86].

In ASIT antihistamines are used not only to relieve symptoms of AR, but also to monitor AE,

induced by administration of therapeutic allergen at the beginning of therapy, being drugs of choice in mild and moderate severity of AE. The use of second generation H1-receptor blockers, not affecting blood pressure indicators and not inhibiting the respiratory centre, is shown. Severe AE may require administration of inhaled and systemic steroids, bronchodilators and epinephrine.

There is no data on negative impact of antihistamines on ASIT efficacy. According to the meta-analysis of 2021, the use of antihistamines as premedication during ASIT significantly reduces the risk of moderate and severe systemic responses and increases the chances of reaching the target maintenance dose of the allergen [87].

Omalizumab allows to reduce to the minimum the incidence of anaphylactic reactions while maintaining efficacy of ASIT [88].

Thus, ASIT is possible with the use of drugs of basic therapy.

ASIT is currently recognized as the only clinically effective, disease-modifying way to treat IgE-mediated allergic diseases (rhinitis, asthma, food allergy), providing the achievement of a long-term effect, ongoing after discontinuation of treatment [8].

In pediatric practice ASIT, according to some reports, reduces the incidence of new sensitization and transformation of rhinitis in asthma.

Specific immunotherapy is most studied and applicable in actual practice in disease, caused by pollen of plants and house dust mites.

REFERENCES/ ЛИТЕРАТУРА

1. Noon L. Prophylactic inoculations against hay fever. *Lancet*. 1911; 1: 1572–1573.
2. Cooke RA. The treatment of hay fever by active immunization. *Laryngoscope*. 1914; 25: 108–112.
3. Freeman J. Further observations on the treatment of hay fever by hypodermic inoculations of pollen vaccine. *Lancet*. 1911; 178: 814–817.
4. Meyer-Pittroff R, Behrendt H, Ring J. Specific immuno-modulation and therapy by means of high pressure treated allergens. *High Pressure Research*. 2007; 27 (1): 63–67.
5. Воробьева ОВ, Гушчин ИС. Исторические предпосылки создания метода аллерген-специфической иммунотерапии (К 100-летию открытия). *Российский аллергологический журнал*. 2010; 5: 17–28. [Vorobjeva OV, Gushchin IS. Istoricheskie predposylki sozdaniya metoda allergen-specificheskoy immunoterapii (K 100-letiyu otkrytiya). *Rossijskij allergologicheskij zhurnal*. 2010; 5: 17–28. (In Russ.)]
6. Гушчин ИС, Курбачева ОМ. Аллергия и аллерген-специфическая иммунотерапия. М.: Фармарус Принт Медиа. 2010; 228: 16. [Gushchin IS, Kurbacheva OM. *Allergiya i allergen-specificheskaya immunoterapiya*. М.: Farmarus Print Media. 2010; 228: 16. (In Russ.)]
7. Sahiner UM, Giovannini M, Escribese MM et al. Mechanisms of Allergen Immunotherapy and Potential Biomarkers for Clinical Evaluation. *J Pers Med*. 2023; 13 (5): 845. <https://doi.org/10.3390/jpm13050845>.
8. Mitsias D, Xepapadaki P, Makris M, Papadopoulos N. Immunotherapy in allergic diseases — improved understanding and innovation for enhanced effectiveness. *Curr Opin Immunol*. 2020; 66: 1–8. <https://doi.org/10.1016/j.coi.2020.02.005>.

9. Вишнева ЕА, Намазова-Баранова ЛС, Алексеева АА и др. Аллерген-специфическая иммунотерапия у детей: современное состояние вопроса. Педиатрическая фармакология. 2016; 13 (4): 404–408. [Vishneva EA, Namazova-Baranova LS, Alekseeva AA et al. Allergen-spetsificheskaya immunoterapiya u detei: sovremennoe sostoyanie voprosa. *Pediatricheskaya farmakologiya*. 2016; 13 (4): 404–408. (In Russ.)]
10. Shamji MH, Durham SR. Mechanisms of allergen immunotherapy for inhaled allergens and predictive biomarkers. *J. Allergy Clin. Immunol.* 2017; 140 (6): 1485–1498. <https://doi.org/10.1016/j.jaci.2017.10.010>.
11. Alvaro-Lozano M, Akdis CA, Akdis M et al. EAACI Allergen Immunotherapy User's Guide. *Pediatr Allergy Immunol.* 2020; 25 (25): 1–101. <https://doi.org/10.1111/pai.13189>.
12. Pavón-Romero GF, Parra-Vargas MI, Ramírez-Jiménez F et al. Allergen Immunotherapy: Current and Future Trends. *Cells.* 2022; 11 (2): 212. <https://doi.org/10.3390/cells11020212>.
13. Yu JC, Khodadadi H, Malik A et al. Innate Immunity of Neonates and Infants. *Front Immunol.* 2018; 9: 1759. <https://doi.org/10.3389/fimmu.2018.01759>.
14. Belderbos M, Levy O, Bont L. Neonatal innate immunity in allergy development. *Curr Opin Pediatr.* 2009; 21 (6): 762–769. <https://doi.org/10.1097/MOP.0b013e3283325e3a>.
15. Zielen S, Devillier P, Heinrich J et al. Sublingual immunotherapy provides long-term relief in allergic rhinitis and reduces the risk of asthma: A retrospective, real-world database analysis. *Allergy.* 2018; 73 (1): 165–177. <https://doi.org/10.1111/all.13213>.
16. Матвеева ЛП, Ермакова ЛА. Экспертиза лечения поллиноза у детей аллергоидами и аллергенами. Проблемы экспертизы в медицине. 2006; 3: 42–44. [Matveeva LP, Ermakova LA. Ekspertiza lecheniya pollinoza u detej allergoidami i allergenami. *Problemy ekspertizy v medicine*. 2006; 3: 42–44. (In Russ.)]
17. Barnes C, Portnoy JM, Ciccio CE, Pacheco F. A comparison of subject room dust with home vacuum dust for evaluation of dust-borne aeroallergens. *Ann Allergy Asthma Immunol.* 2013; 110 (5): 375–379. <https://doi.org/10.1016/j.anai.2013.02.010>.
18. Mattsson L, Lundgren T, Everberg H et al. Prostatic kallikrein: a new major dog allergen. *J Allergy Clin Immunol.* 2009; 123 (2): 362–368. <https://doi.org/10.1016/j.jaci.2008.11.021>.
19. Van Ree R, Chapman MD, Ferreira F, et al. The CREATE Project: Development of certified reference materials for allergenic products and validation of methods for their quantification. *Allergy: European Journal of Allergy and Clinical Immunology.* 2008; 63 (3): 310–326. <https://doi.org/10.1111/j.1398-9995.2007.01612.x>.
20. Larenas-Linnemann D, Cox LS. Immunotherapy and Allergy Diagnostics Committee of the American Academy of Allergy, Asthma and Immunology. European allergen extract units and potency: review of available information. *Ann Allergy Asthma Immunol.* 2008; 100 (2): 137–145. [https://doi.org/10.1016/S1081-1206\(10\)60422-X](https://doi.org/10.1016/S1081-1206(10)60422-X).
21. Государственная фармакопея. XIII изд. ОФС.1.7.1.0001.15. Аллергены. Веб-ресурс: <https://pharmacopoeia.ru/ofs-1-7-1-0001-15-allergeny/#%D0%98%D0%A1%D0%9F%D0%AB%D0%A2%D0%90%D0%9D%D0%98%D0%AF>. Время доступа 08.06.2023. [Gosudarstvennaya farmakopeya. XIII izd. OFS.1.7.1.0001.15. Allergeny. Veb-resurs: <https://pharmacopoeia.ru/ofs-1-7-1-0001-15-allergeny/#%D0%98%D0%A1%D0%9F%D0%AB%D0%A2%D0%90%D0%9D%D0%98%D0%AF>. Vremya dostupa 08.06.2023. (In Russ.)]
22. Lund K, Kito H, Skydtsgaard MB et al. The Importance of Tablet Formulation on Allergen Release Kinetics and Efficiency: Comparison of Freeze-dried and Compressed Grass Pollen Sublingual Allergy Immunotherapy Tablet Formulations. *Clin Ther.* 2019; 41 (4): 742–753. <https://doi.org/10.1016/j.clinthera.2019.02.008>.
23. Wahn U, Bachert C, Heinrich J et al. Real-world benefits of allergen immunotherapy for birch pollen-associated allergic rhinitis and asthma. *Allergy.* 2019; 74 (3): 594–604. <https://doi.org/10.1111/all.13598>.
24. Gangl K, Niederberger V, Valenta R. Multiple grass mixes as opposed to single grasses for allergen immunotherapy in allergic rhinitis. *Clinical and Experimental Allergy: Journal of the British Society for Allergy and Clinical Immunology.* 2013; 43 (11): 1202–1216. <https://doi.org/10.1111/cea.12128>.
25. Gunawardana NC, Durham SR. New approaches to allergen immunotherapy. *Ann Allergy Asthma Immunol.* 2018; 121 (3): 293–305. <https://doi.org/10.1016/j.anai.2018.07.014>.
26. Ring J, Guterthum J. 100 years of hyposensitization: history of allergen-specific immunotherapy (ASIT). *Allergy.* 2011; 66 (6): 713–724. <https://doi.org/10.1111/j.1398-9995.2010.02541.x>.
27. Valenta R, Niederberger V. Recombinant allergens for immunotherapy. *J Allergy Clin Immunol.* 2007; 119: 826–830.
28. Riabova K, Karsonova AV, van Hage M, et al. Molecular Allergen-Specific IgE Recognition Profiles and Cumulative Specific IgE Levels Associated with Phenotypes of Cat Allergy. *Int J Mol Sci.* 2022; 23 (13): 6984. Published 2022 Jun 23. <https://doi.org/10.3390/ijms23136984>.

29. Ding B, Lai Y, Lu Y. Combined application of dupilumab and mite allergen-specific immunotherapy in children with moderate to severe atopic dermatitis. *Allergol Immunopathol (Madr)*. 2023; 51 (2): 184–190. Published 2023 Mar 1. <https://doi.org/10.15586/aei.v51i2.778>.
30. Casale TB, Busse WW, Kline JN et al. Omalizumab pretreatment decreases acute reactions after rush immunotherapy for ragweed-induced seasonal allergic rhinitis. *J Allergy Clin Immunol*. 2006; 117: 134–140.
31. Rodríguez Del Río P, Vidal C, Just J, et al. The European Survey on Adverse Systemic Reactions in Allergen Immunotherapy (EASSI): A paediatric assessment. *Pediatr Allergy Immunol*. 2017; 28 (1): 60–70. <https://doi.org/10.1111/pai.12660>.
32. Calderón MA, Vidal C, Rodríguez Del Río P, et al. European Survey on Adverse Systemic Reactions in Allergen Immunotherapy (EASSI): a real-life clinical assessment. *Allergy*. 2017; 72 (3): 462–472. <https://doi.org/10.1111/all.13066>.
33. Calderón MA, Simons FE, Malling HJ, et al. Sublingual allergen immunotherapy: mode of action and its relationship with the safety profile. *Allergy*. 2012; 67 (3): 302–311. <https://doi.org/10.1111/j.1398-9995.2011.02761.x>.
34. Di Bona D, Magistà S, Masciopinto L, et al. Safety and treatment compliance of subcutaneous immunotherapy: A 30-year retrospective study. *Respir Med*. 2020; 161: 105843. <https://doi.org/10.1016/j.rmed.2019.105843>.
35. Pfaar O, Bachert C, Bufe A, et al. Guideline on allergen-specific immunotherapy in IgE-mediated allergic diseases: S2k Guideline of the German Society for Allergology and Clinical Immunology (DGAKI), the Society for Pediatric Allergy and Environmental Medicine (GPA), the Medical Association of German Allergologists (AeDA), the Austrian Society for Allergy and Immunology (ÖGAI), the Swiss Society for Allergy and Immunology (SGAI), the German Society of Dermatology (DDG), the German Society of Oto- Rhino- Laryngology, Head and Neck Surgery (DGHNO-KHC), the German Society of Pediatrics and Adolescent Medicine (DGKJ), the Society for Pediatric Pneumology (GPP), the German Respiratory Society (DGP), the German Association of ENT Surgeons (BV-HNO), the Professional Federation of Paediatricians and Youth Doctors (BVKJ), the Federal Association of Pulmonologists (BDP) and the German Dermatologists Association (BVDD). *Allergo J Int*. 2014; 23 (8): 282–319. <https://doi.org/10.1007/s40629-014-0032-2>.
36. Sánchez-Borges M, Bernstein DI, Calabria C. Subcutaneous Immunotherapy Safety: Incidence per Surveys and Risk Factors. *Immunol Allergy Clin North Am*. 2020; 40 (1): 25–39. <https://doi.org/10.1016/j.iac.2019.09.001>.
37. Roberts G, Pfaar O, Akdis CA, et al. EAACI Guidelines on Allergen Immunotherapy: Allergic rhinoconjunctivitis. *Allergy*. 2018; 73 (4): 765–798. <https://doi.org/10.1111/all.13317>.
38. Cox L, Nelson H, Lockey R, et al. Allergen immunotherapy: a practice parameter third update. *J Allergy Clin Immunol*. 2011; 127 (1): 1–55. <https://doi.org/10.1016/j.jaci.2010.09.034>.
39. Pitsios C, Demoly P, Bilò MB, et al. Clinical contraindications to allergen immunotherapy: an EAACI position paper. *Allergy*. 2015; 70 (8): 897–909. <https://doi.org/10.1111/all.12638>.
40. Pitsios C, Tsoumani M., Bilò MB, et al. Contraindications to immunotherapy: a global approach. *Clin Transl Allergy*. 2019; 9: 45. <https://doi.org/10.1186/s13601-019-0285-4>.
41. Bousquet J, Pfaar O, Togias A, et al. 2019 ARIA Care pathways for allergen immunotherapy. *Allergy*. 2019; 74 (11): 2087–2102. <https://doi.org/10.1111/all.13805>.
42. Agache I, Lau S, Akdis CA, et al. EAACI Guidelines on Allergen Immunotherapy: House dust mite-driven allergic asthma. *Allergy*. 2019; 74 (5): 855–873. <https://doi.org/10.1111/all.13749>.
43. Pfaar O, Ankermann T, Augustin M, et al. Guideline on allergen immunotherapy in IgE-mediated allergic diseases: S2K Guideline of the German Society of Allergology and Clinical Immunology (DGAKI), Society of Pediatric Allergology and Environmental Medicine (GPA), Medical Association of German Allergologists (AeDA), Austrian Society of Allergology and Immunology (ÖGAI), Swiss Society for Allergology and Immunology (SSAI), German Dermatological Society (DDG), German Society of Oto-Rhino-Laryngology, Head and Neck Surgery (DGHNO-KHC), German Society of Pediatrics and Adolescent Medicine (DGKJ), Society of Pediatric Pulmonology (GPP), German Respiratory Society (DGP), German Professional Association of Otolaryngologists (BVHNO), German Association of Paediatric and Adolescent Care Specialists (BVKJ), Federal Association of Pneumologists, Sleep and Respiratory Physicians (BdP), Professional Association of German Dermatologists (BVDD). *Allergol Select*. 2022; 6: 67–232. <https://doi.org/10.5414/ALX02331E>.
44. Ferrando M, Racca F, Madeira LNG, et al. A critical appraisal on AIT in childhood asthma. *Clin Mol Allergy*. 2018; 16: 6. <https://doi.org/10.1186/s12948-018-0085-8>.
45. Cox L, Larenas-Linnemann D, Lockey RF, Passalacqua G. Speaking the same language: The World Allergy Organization Subcutaneous Immunotherapy Systemic Reaction Grading System. *J Allergy Clin Immunol*. 2010; 125 (3): 569–574, 574.e1–574.e7. <https://doi.org/10.1016/j.jaci.2009.10.060>.

46. Iemoli E, Borgonovo L, Fusi A, et al. Sublingual allergen immunotherapy in HIV-positive patients. *Allergy*. 2016; 71: 412–415. <https://doi.org/10.1111/all.12713>.
47. Linneberg A, Madsen F, Skaaby T. Allergen-specific immunotherapy and risk of autoimmune disease. *Curr Opin Allergy Clin Immunol*. 2012; 12 (6): 635–639. <https://doi.org/10.1097/ACI.0b013e3283588c8d>.
48. Wollenberg A, Barbarot S, Bieber T, et al. European Dermatology Forum (EDF), the European Academy of Dermatology and Venereology (EADV), the European Academy of Allergy and Clinical Immunology (EAACI), the European Task Force on Atopic Dermatitis (ETFAD), European Federation of Allergy and Airways Diseases Patients' Associations (EFA), the European Society for Dermatology and Psychiatry (ESDaP), the European Society of Pediatric Dermatology (ESPD), Global Allergy and Asthma European Network (GA2LEN) and the European Union of Medical Specialists (UEMS). Consensus-based European guidelines for treatment of atopic eczema (atopic dermatitis) in adults and children: part II. *J Eur Acad Dermatol Venereol*. 2018; 32 (6): 850–878. <https://doi.org/10.1111/jdv.14888>.
49. Egan M, Atkins D. What Is the Relationship Between Eosinophilic Esophagitis (EoE) and Aeroallergens? Implications for Allergen Immunotherapy. *Curr Allergy Asthma Rep*. 2018; 18 (8): 43. <https://doi.org/10.1007/s11882-018-0798-2>.
50. Wang C, Bao Y, Chen J, et al. Chinese Society of Allergy (CSA) and Chinese Allergic Rhinitis Collaborative Research Group (C2AR2G). Chinese Guideline on Allergen Immunotherapy for Allergic Rhinitis: The 2022 Update. *Allergy Asthma Immunol Res*. 2022; 14 (6): 604–652. <https://doi.org/10.4168/aaair.2022.14.6.604>.
51. Gao Y, Lin X, Ma J, et al. Enhanced Efficacy of Dust Mite Sublingual Immunotherapy in Low-Response Allergic Rhinitis Patients after Dose Increment at 6 Months: A Prospective Study. *International archives of allergy and immunology*. 2020; 181 (4): 311–319. <https://doi.org/10.1159/000505746>.
52. Chen S, Zheng Y, Chen B, et al. Clinical Response to Subcutaneous *Dermatophagoides pteronyssinus* Immunotherapy in Children with Allergic Rhinitis and Asthma Is Independent of Sensitization to *Blomia tropicalis* Allergens. *Int Arch Allergy Immunol*. 2019; 178 (2): 201–210. <https://doi.org/10.1159/000494389>.
53. Fortescue R, Kew KM, Leung MST. Sublingual immunotherapy for asthma. *Cochrane Database Syst Rev*. 2020; 9: CD011293. <https://doi.org/10.1002/14651858.CD011293.pub3>.
54. Radulovic S, Calderon MA, Wilson D, Durham S. Sublingual immunotherapy for allergic rhinitis. *Cochrane Database of Systematic Reviews*. 2010; (12). <https://doi.org/10.1002/14651858.CD002893.pub2>.
55. Клинические рекомендации. Атопический дерматит. 2021. https://cr.minzdrav.gov.ru/recomend/265_2. Дата обращения 18.06.2023. [Klinicheskie rekomendacii. Atopicheskij dermatit. 2021. https://cr.minzdrav.gov.ru/recomend/265_2. Vremya dostupa 18.06.2023 (In Russ.)]
56. Lee J, Lee H, Noh S, et al. Retrospective Analysis on the Effects of House Dust Mite Specific Immunotherapy for More Than 3 Years in Atopic Dermatitis. *Yonsei Med J*. 2016; 57 (2): 393–398. <https://doi.org/10.3349/ymj.2016.57.2.393>.
57. Liu L, Chen J, Xu J, et al. Sublingual immunotherapy of atopic dermatitis in mite-sensitized patients: a multi-centre, randomized, double-blind, placebo-controlled study. *Artif Cells Nanomed Biotechnol*. 2019; 47 (1): 3540–3547. <https://doi.org/10.1080/21691401.2019.1640709>.
58. Marogna M, Spadolini I, Massolo A, et al. Long-lasting effects of sublingual immunotherapy according to its duration: A 15-year prospective study. *Journal of Allergy and Clinical Immunology*. 2010; 126 (5): 969–975. <https://doi.org/10.1016/j.jaci.2010.08.030>.
59. Trebuchon F, Lhéritier-Barrand M, David M, Demoly P. Characteristics and management of sublingual allergen immunotherapy in children with allergic rhinitis and asthma induced by house dust mite allergens. *Clin Transl Allergy*. 2014; 4: 15. <https://doi.org/10.1186/2045-7022-4-15>.
60. Demoly P, Meziane L, LeGall M, et al. Safety and tolerability of house dust mite tablets in sublingual immunotherapy. *J Allergy Clin Immunol*. 2008; 121: S128.
61. Mosbech H, Deckelmann R, de Blay F, et al. Standardized quality (SQ) house dust mite sublingual immunotherapy tablet (ALK) reduces inhaled corticosteroid use while maintaining asthma control: a randomized, double-blind, placebo-controlled trial. *J Allergy Clin Immunol*. 2014; 134: 568–575.
62. Mösges R, Ritter B, Kayoko G, Passali D, Allekotte S. Carbamylated monomeric allergoids as a therapeutic option for sublingual immunotherapy of dust mite- and grass pollen-induced allergic rhinoconjunctivitis: a systematic review of published trials with a meta-analysis of treatment using Lais tablets. *Acta Dermatovenereol Alp Pannonica Adriat*. 2010 Oct; 19 (3): 3–10.
63. Abramson MJ, Puy RM, Weiner JM. Injection allergen immunotherapy for asthma. *Cochrane Airways Group, ed. Cochrane Database of Systematic Reviews*. Published on line August 4, 2010. <https://doi.org/10.1002/14651858.CD001186.pub2>.

64. Гайдук ИМ, Макарова ИВ, Трусова ОВ, Брейкин ДВ, Сухорукова ВГ, Аракелян РН, Коростовцев ДС. Аллерген-специфическая иммунотерапия пыльцевыми аллергоидами у детей. Российский аллергологический журнал. 2009; 1: 45–50. [Gajduk IM, Makarova IV, Trusova OV, Brejkin DV, Suhorukova VG, Arakelyan RN, Korostovtsev DS. Allergen-specificeskaya immunoterapiya pyl'cevyimi allergoidami u detej. Rossijskij allergologicheskij zhurnal. 2009; 1: 45–50. (In Russ.)]
65. Worm M, Rak S, DeBlay F et al. Sustained efficacy and safety of a 300IR daily dose of a sublingual solution of birch pollen allergen extract in adults with allergic rhinoconjunctivitis: result of a double-blind, placebo-controlled study. *Clinical and Translational Allergy*. 2014; 4: 1–11.
66. Wahn U, Tabar A, Kuna P et al. Efficacy and safety of 5-grass-pollen sublingual immunotherapy tablets in pediatric allergic rhinoconjunctivitis. *Journal of Allergy and Clinical Immunology*. 2009; 123 (1): 160–166.e3. <https://doi.org/10.1016/j.jaci.2008.10.009>.
67. Valovirta E, Petersen TH, Piotrowska T, et al. Results from the 5-year SQ grass sublingual immunotherapy tablet asthma prevention (GAP) trial in children with grass-pollen allergy. *J Allergy Clin Immunol*. 2018; 141 (2): 529–538.e13. <https://doi.org/10.1016/j.jaci.2017.06.014>.
68. Nolte H, Bernstein DI, Nelson HS, et al. Efficacy and Safety of Ragweed SLIT-Tablet in Children with Allergic Rhinoconjunctivitis in a Randomized, Placebo-Controlled Trial. *J Allergy Clin Immunol Pract*. 2020; 8 (7): 2322–2331.e5. <https://doi.org/10.1016/j.jaip.2020.03.041>.
69. Maloney J, Berman G, Gagnon R, et al. Sequential Treatment Initiation with Timothy Grass and Ragweed Sublingual Immunotherapy Tablets Followed by Simultaneous Treatment Is Well Tolerated. *J Allergy Clin Immunol Pract*. 2016; 4 (2): 301–309.e2. <https://doi.org/10.1016/j.jaip.2015.11.004>.
70. Bousquet J, Hejjaoui A, Dhivert H, et al. Immunotherapy with a standardized *Dermatophagoides pteronyssinus* extract. Systemic reactions during the rush protocol in patients suffering from asthma. *J Allergy Clin Immunol*. 1989; 83 (4): 797–802. [https://doi.org/10.1016/0091-6749\(89\)90017-1](https://doi.org/10.1016/0091-6749(89)90017-1).
71. Смолкин ЮС, Максимова АВ. Особенности профилактической вакцинации детей с аллергическими заболеваниями. Учеб.-метод. пособие. Москва. Изд-во Академии постдипломного образования ФГБУ ФНКЦ ФМБА России, 2020. 52 с. ISBN 978-5-7151-0559-2. [Smolkin YUS, Maksimova AV. Osobennosti profilakticheskoy vakcinacii detej s allergicheskimi zabolevaniyami. Ucheb.-metod. posobie. Moskva. Izd-vo Akademii postdiplomnogo obrazovaniya FGBU FNKC FMBA Rossii, 2020. 52 s. ISBN 978-5-7151-0559-2. (In Russ.)]
72. Chen K-W, Ziegelmayer P, Ziegelmayer R et al. Selection of house dust mite-allergic patients by molecular diagnosis may enhance success of specific immunotherapy. *Journal of Allergy and Clinical Immunology*. 2019; 143 (3): 1248–1252.
73. Sastre J, Rodriguez F, Campo P, et al. Adverse reactions to immunotherapy are associated with different patterns of sensitization to grass allergens. *Allergy*. 2015; 70 (5): 598–600.
74. Shamji MH, Kappen JH, Akdis M, et al. Biomarkers for monitoring clinical efficacy of allergen immunotherapy for allergic rhinoconjunctivitis and allergic asthma: an EAACI Position Paper. *Allergy*. 2017; 72 (8): 1156–1173. <https://doi.org/10.1111/all.13138>.
75. Барычева ЛЮ, Душина ЛВ, Медведенко ЮН. Изменение реактивности базофилов и синтеза специфических иммуноглобулинов Е под влиянием аллерген-иммунотерапии. *Аллергология и иммунология в педиатрии*. 2020; 1 (64): 15–23. <https://doi.org/10.24412/2500-1175-2021-1-15-23>. [Barycheva LYU, Dushina LV, Medvedenko YUN. Izmenenie reaktivnosti bazofilov i sinteza specificheskikh immunoglobulinov E pod vliyaniem allergen-immunoterapii. *Allergologiya i immunologiya v pediatrii*. 2020; 1 (64): 15–23. <https://doi.org/10.24412/2500-1175-2021-1-15-23>. (In Russ.)]
76. Частная аллергология. Под ред. Адо А. Д. М.: «Медицина», 1976. 512 с. [Chastnaya allergologiya. Pod red. Ado A. D. M.: «Medicina», 1976. 512 s. (In Russ.)]
77. Pfaar O, Demoly P, Gerth van Wijk R, et al. Recommendations for the standardization of clinical outcomes used in allergen immunotherapy trials for allergic rhinoconjunctivitis: an EAACI Position Paper. *Allergy*. 2014; 69 (7): 854–867.
78. Тактика диагностики и лечения аллергических заболеваний и иммунодефицитов: практическое руководство / под ред. Р. М. Хаитова. — М.: ГЭОТАР-Медиа, 2019. — 152 с. DOI: 10.33029/9704-5200-4-TER-2019-1-152. [Taktika diagnostiki i lecheniya allergicheskikh zabolevanij i immunodeficitov: prakticheskoe rukovodstvo / pod red. R. M. Haitova. — М.: GEOTAR-Media, 2019. — 152 s. DOI: 10.33029/9704-5200-4-TER-2019-1-152. (In Russ.)]
79. Bae JM, Choi YY, Park CO, et al. Efficacy of allergen-specific immunotherapy for atopic dermatitis: a systematic review and meta-analysis of randomized controlled trials. *J Allergy Clin Immunol*. 2013; 132: 110–117. <https://doi.org/10.1016/j.jaci.2013.02.044>.
80. Traidl S, Werfel T. Allergen immunotherapy for atopic dermatitis. *Hautarzt*. 2021; 72 (12): 1103–1112. <https://doi.org/10.1007/s00105-021-04909-y>.

81. Valera CP, Coelho EB, Galvao CES, et al. Efficacy of House Dust Mite Sublingual Immunotherapy in Patients with Atopic Dermatitis: A Randomized, Double-Blind, Placebo-Controlled Trial Randomized Controlled Trial. *J Allergy Clin Immunol Pract.* 2022; 10 (2): 539–549.e7. <https://doi.org/10.1016/j.jaip.2021.10.060>.
82. Enrique E, Pineda F, Malek T, et al. Sublingual immunotherapy for hazelnut food allergy: a randomized, double-blind, placebo-controlled study with a standardized hazelnut extract. *J Allergy Clin Immunol.* 2005; 116 (5): 1073–1079. <https://doi.org/10.1016/j.jaci.2005.08.027>.
83. Romantsik O, Tosca MA, Zappettini S, Calevo MG. Oral and sublingual immunotherapy for egg allergy (Review). *Cochrane Database Syst Rev.* 2018; (4): CD010638.
84. Pajno GB, Fernandez-Rivas M, Arasi S, et al. EAACI guidelines on allergen immunotherapy: IgE mediated food allergy. *Allergy.* 2018; 73: 799–815.
85. AAAAI current status of oral immunotherapy (ICU) for the treatment of food allergies. [accessed April 1, 2020]; available on the Internet: <https://www.aaaai.org/conditions-and-treatments/library/allergy-library/oit>.
86. Virchow JC, Backer V, Kuna P, et al. Efficacy of a House Dust Mite Sublingual Allergen Immunotherapy Tablet in Adults With Allergic Asthma. *JAMA.* 2016; 315 (16): 1715. <https://doi.org/10.1001/jama.2016.3964>.
87. Wang L, Wang C, Lou H, et al. Antihistamine premedication improves safety and efficacy of allergen immunotherapy. *Ann Allergy Asthma Immunol.* 2021; 127 (3): 363–371.
88. Lee TH, Chan JKC, Lau PC, et al. Peanut allergy and oral immunotherapy. *Hong Kong Med J.* 2019; 25 (3): 228–234. <https://doi.org/10.12809/hkmj187743>.

THE AUTHORS' CONTRIBUTION TO THE WORK

Yuri S. Smolkin — publication design development, review of the article critical content, manuscript text editing.

Olga V. Trusova — publication design development, review of publications on the article subject, manuscript text writing and editing, review of the article critical content.

Zuleikha A. Aliskandieva, Liudmila Y. Barycheva, Alexey D. Bogomazov, Ksenia A. Bocharova, Yuliya N. Emelina, Andrey V. Kamaev, Inna A. Larkova, Aishat Z. Markhaichuk, Sergey S. Masalskiy, Natalia B. Migacheva, Aleksandr S. Prilutskiy, Elena V. Stezhkina, Rezeda M. Fayzullina, Rezeda F. Khakimova, Ella V. Churyukina, Natal'ya V. Shakhova, Tatiyana V. Shilova — review of publications on the article subject, manuscript text writing and editing.

ВКЛАД АВТОРОВ В РАБОТУ

Смолкин Ю. С. — разработка дизайна публикации, проверка критически важного содержания статьи, редактирование текста рукописи.

Трусова О. В. — разработка дизайна публикации, обзор публикаций по теме статьи, написание и редактирование текста рукописи, проверка критически важного содержания статьи.

Алискандиева З. А., Барычева Л. Ю., Богомазов А. Д., Бочарова К. А., Емелина Ю. Н., Камаев А. В., Ларькова И. А., Мархайчук А. З., Масальский С. С., Мигачева Н. Б., Прилуцкий А. С., Стежкина Е. В., Файзуллина Р. М., Хакимова Р. Ф., Чурюкина Э. В., Шахова Н. В., Шилова Т. В. — обзор публикаций по теме статьи, написание и редактирование текста рукописи.